Intraocular pressure control of a novel glaucoma drainage device - *in vitro* and *in vivo* studies

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Abstract

- **AIM:** To evaluate the intraocular pressure (IOP) control of an artificial trabeculum drainage system (ATDS), a newly designed glaucoma drainage device, and postoperative complications in normal rabbit eyes.
- **METHODS:** Pressure drops in air and fluid of 30 ATDS were measured after being connected to a closed manometric system. Twenty of them were then chosen and implanted randomly into the eyes of 20 rabbits. Postoperative slit-lamp, gonioscopic examination and IOP measurements were recorded periodically. Ultrasound biomicroscopy and B-scan ultrasonography were also used to observe the complications. Eyes were enucleated on day 60.
- **RESULTS:** Pressure drops of 4.6-9.4 mm Hg were obtained at physiological aqueous flow rates in the tests *in vitro*. The average postoperative IOP of the experimental eyes (11.6-12.8 mm Hg) was lower than the controls significantly (P<0.05) at each time point. Complications of hemorrhage (n=1), cellulosic exudation (two cases) and local iris congestion (two cases) were observed. The lumina of the devices were devoid of obstructions in all specimens examined and a thin fibrous capsule was found around the endplate.
- **CONCLUSION:** ATDS reduce IOP effectively. However, further studies on the structure are needed to reduce complications.

- **KEYWORDS:** drainage device; aqueous humor; outflow; intraocular pressure; rabbit

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INTRODUCTION

Glaucoma is the second most common cause of blindness and the leading cause of irreversible blindness[1]. Glaucoma filtration surgery is a fistulizing procedure that provides an alternative drainage route allowing aqueous to escape from the anterior chamber (AC) to the subconjunctiva in order to lower intraocular pressure (IOP), which includes trabeculectomy and drainage implant surgery. Implantation of glaucoma drainage devices (GDDs) has become a standard procedure in various forms of complicated and refractory glaucoma with comparable IOP control and duration of benefit[2-3]. In certain conditions, such as neovascular glaucoma, irido-corneal syndrome, penetrating keratopathy with glaucoma, and glaucoma following retinal detachment surgery, uveitis, or trauma and so on, it is becoming the primary operation[4]. Such devices enable percolation of aqueous liquid through a tube to a filtering plate in the subconjunctival space, to the Schlemm’s canal, or to the suprachoroidal space[5].

There have been many contemporary GDDs commercially available[6] since Molteno implant was invented in 1969[7], but success rates of most clinical series were not satisfied[8-9]. There are a number of unsolved clinical drawbacks of existing systems[10]. While partly attributable to the complicated manifestations and strong wound-healing trend of cases typically selected for implantation, various complications also lead to filtration failure[11-14]. The most significant complication related to exposure is endophthalmitis. Others include hypotony, shallow AC, choroidal effusion, suprachoroidal hemorrhage, tube migration and tube obstruction. The origin of most complications can be traced to design inadequacies, poor flow control, lack of set resistance and suboptimal material biocompatibility[3].

In general, there are 2 types of GDDs[3,4,6,15], with or without set resistance mechanism or pressure sensible valve. The valved implants have a pressure-regulating mechanism to minimize
overdrainage. The Ahmed glaucoma valve\cite{16-17}, in particular, is proved to function as a real valve that closely regulates pressure within a desired range by a variable resistance in response to changes in flow rate. But the potential site for obstruction by inflammatory debris and valve membrane adhesion\cite{18}, especially in Asian eyes\cite{19}, may cause surgery failure. On the other hand, hypotony caused by the bulk outflow of aqueous in the early postoperative period is much common in valveless implants. The inserted tube dimension is too large to produce resistance when aqueous humor flows through it at physiological rates. The Baerveldt GDD is available with a surface area of 350 mm² and requires temporary flow restriction to avoid early postoperative hypotony\cite{20}. Either a two staged procedure or ligature technique\cite{11,21-23}, with or without fenestration of the tube, are therefore required to produce a temporary restriction of flow. However, these methods are sometimes cumbersome and time consuming, and many researchers\cite{23-24} had proved that it was not possible to regulate pressure in a reliable and predictable way merely by constricting the tube lumen.

On the basis of hydrodynamic principles, we developed a new restricted GDD without valve membranes, which is named artificial trabeculum drainage system (ATDS). The main purpose of this study is to evaluate flow characteristics of ATDS, and observe IOP change and complications after implanted in rabbit eyes.

MATERIALS AND METHODS

Artificial Trabeculum Drainage System  Following the concept of tube and plate GDD, ATDS consists of a T-shaped silastic tube (Medical Silicon Rubber Technical Institute of Rubber Goods Design Academy, Beijing, China) and a pear-shaped plate (Institute of Advanced Manufacturing Technology, School of Mechanical Engineering, Xi’an Jiaotong University, Xi’an, China) made of medical-grade polyurethane (PUR) (Figure 1).

The T-tube has the same dimension of 600 µm in outer diameter and 300 µm in inner as the single round tube of GDD available in the market. Several micropores, with diameter of 250 µm, distribute to the 6 mm-long horizontal tube to decrease the blockage of the tube (Figure 2).

The pressure confined system (PCS) on the endplate, which is also a silastic tube, with the inner diameter of 80 µm, circles as a certain mode. This mode is selected from several designs in different tube lengths, calibers and circling ways, by calculating and screening step by step using Poiseuille’s law, Bernoulli’s formula and FLUNT hydrodynamic software. Pressure drop versus flow rate from theoretical calculations is shown as Figure 3. The local pressure impairments caused by 8 angles with different degrees and a diameter change (from 300 µm to 80 µm) are calculated from Bernoulli formula:

\[ h_l = \lambda \frac{1}{d} \sum \zeta \frac{u^2}{2g} \]

But the summation is too small to disturb the linear correlation because of the slow flow.

![Figure 1 Simulated construction of ATDS](image1)

![Figure 2 Structure of T-shaped tube](image2)

![Figure 3 Pressure drops of PCS at different flow rates](image3)
Hydrodynamic Test  The characteristic parameters were tested through a flow rig consisted of a tubing compression pump (Model T-Y, TongYi Inc., Shanghai, China), a bridge amplifier (Model ML110, AD Instruments, New South Wales, Australia), a recorder (Model ML200, AD Instruments, New South Wales, Australia), a pressure transducer (Powerlab, AFR Instruments, Tokyo, Japan) and two three-way locks (Figure 4). Pressure changes were recorded and analyzed using Chart 4 software.

Degassed balanced salt solution (BSS) was infused by the pump with initial flow rates preset to 0.6, 1.2, 2.4, 4.8, 9.0, 15.0, 24.0, 48.0 µL/min. All gas bubbles were flushed out when BSS filled all the system. At this point, the two three-way locks were turned to open the system to atmospheric pressure and the pressure reading was zeroed on the recorder. The first three-way lock was then turned to obtain a closed system and the infusion pump was started. The two pressure readings were taken at the same time, and the difference between them was recorded. Repeated flow measurements were taken (n=3). Pressure drops in air and water of 30 ATDSs were tested respectively. Each device was measured during a 10min interval while a constant flow of fluid was pumped into the system.

Experimental Animals  A prospective, randomized study was performed using 20 male and female New Zealand White rabbits initially weighing 2.5 to 3.0 kg. The experiment was performed in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. The project was also approved by the Local Animal Research Review committee of the First Affiliated Hospital of Xi'an Jiaotong University, Xi'an, China. All animals were maintained in a 12-hour day and 12-hour night cycle. They were fed and had access to water ad libitum.

Twenty ATDSs were implanted into the unilateral eyes of the rabbits. The horizontal part of the T-shaped tube was inserted into the AC while the endplate was placed subconjunctivally posterior to the equator of the eyeball (Figure 1B). The fellow eyes were served as control.

Surgical Procedure  After adequate general anesthesia [3% pentobarbital sodium (1.5-2.0 mL/kg) intravenous injection], the eyes were prepared and draped with sterile towels. The lids were secured with a lid speculum. Topical 0.5% tetracaine hydrochloride was instilled to prevent any discomfort. A pair of eye scissors was used to perform a superotemporal limbal peritomy from the 11- to 3-o’clock meridian. A 1-0 silk retention suture was placed around the superior rectus muscle to hold the eyeball and expose the surgical area. Conjunctiva and fascia were separated from the globe and superficial bleeding vessels over the site of the intended scleral flap were cauterized lightly. A rectangle scleral flap measuring 3 mm×5 mm and of one-half scleral thickness was dissected up to the limbal zone from the 1- to 2-o’clock meridian (Figure 5A). A sterile ATDS was put into the subconjunctival space in terms of sliding the plate along the scleral surface. Adequate tube was kept for next step of insertion (Figure 5B). A 2-mm limbal incision was made with a 45° blade. The horizontal tube was folded together by a toothed forceps parallel to the iris plane, and sent into the AC through the limbal incision. It stretched quickly and returned to the original shape under its natural flexibility (Figure 5C). The scleral flap was closed with 4 interrupted 10-0 nylon sutures, 1 each on the sides and apexes. After that, the T-shaped tube was pulled back along the surface of the eyeball to make sure the horizontal tube was close to the anterior chamber angle. Fix the endplate to sclera with 2 interrupted 8-0 nylon sutures through the two semicircle protrusions on the head of the plate (Figure 5D).
conjunctival flap was sutured to the limbus with 10-0 nylon suture. Tobradex eyedrop and 0.5% erythromycin ointment were applied into conjunctival sac.

All the surgeries were aseptic and performed by the first author. Tobradex eyedrop was instilled thrice daily for 3d.

**Clinical Observation** Postoperative evaluations were performed on postoperative days 1-3, 7, 14, 21, 30 and 60, or more frequent when necessary, consisted of general health, Seidal test and red reflex, IOP measurement by Perkins handheld applanation tonometer (HL-2, Kowa, Japan), anterior segments observation by slit-lamp biomicroscopy (SL-8Z, Topcon, Japan) and ultrasound biomicroscopy (UBM, 840, Humphrey, Germany) with a 50 MHz transducer, vitreo-retinal complications by B-scan ultrasonography (SW-2100, Suowei, China) with a 10 MHz probe onto the eyelid after performing methylcellulose. IOP of bilateral eyes at each time point were evaluated using a paired, 2-tailed, Student’s t-test.

Animals were sacrificed at the end of research (sedation with a lethal dose of intravenous pentobarbital sodium). The ATDS-implanted eyes were enucleated, with care taken not to disturb the tissues around the implant. The appearance of fibrous capsule and the tube lumen were observed.

**RESULTS**

**Hydrodynamic Tests** The pressure drops examined at different flow rates correlated closely with that predicted by Poiseuille’s formula and Bernoulli’s equation (Figure 6). But there was a tendency of larger variation with flow rate increasing.

**General State and Intraocular Pressure** There was no discharge of all the participants, and red reflex remained normal. There was no significant difference between bilateral baseline IOPs (Table 1), and IOPs of the ATDS-implanted eyes were lower than the controls at each time point ($P=0.000$). All the eyes were devoid of hypotony. Seidal tests were negative. The variation of bilateral IOPs is shown as Figure 7.

**Tissue Responses** Illustrated as Figure 8, the thin, lucent and diffused bleb appeared on postoperative days 1 and 3, and it became localized with time. Conjunctival hyperemia triggered by the surgery occurred on day 1 and lightened prominently on day 3 in most of the ATDS-implanted eyes. Cellulosic exudation happened in 2 cases and resolved spontaneously 5 and 7d after implantation. One case of AC hemorrhage occurred on postoperative day 2 (Figure 9). It localized at the tube site, accompanied with moderate corneal edema. The blood was absorbed 1wk after surgery, but corneal edema still existed. Slight iris eversion with actiniform vessels injection was found at the tube site in several eyes (11/20). The local iris congestion was slight and resolved in the end, but two of them aggravated at first, and reached its apex as diffuse crimson red on postoperative days 14 and 25. Cornea edema (8/20) was localized at the site of surgery, and 6 of them were accompanied with iris vessel injection. They were resolved 3-15d after implantation. Tube erosion was found in 2 eyes at nearly the end of observations, but the subconjunctival wound healed well and peritubular filtration was not found. No tube migration or extrusion happened in the experimental eyes. No focal thickening or adherence of cornea and iris was found in the ATDS-implanted eyes in UBM detection. The horizontal part of T-shaped tube was close to the angle of AC (Figure 10A), while the perpendicular part lying straight on the surface of the sclera (Figure 10B). On each time point,
no obstruction was found in tube lumen of all the samples. There was a low echogenic space between bleb tissue and globe, with small punctiform echogenic distribution in the B-scan ultrasonography image (Figure 10C). No ciliary body detachment, suprachoroidal hemorrhage or retinal detachment was found according to the ultrasonography detections.

Enucleated Eyes The local tissue reaction typically consisted of a pink, thin and tenacious capsule. This smooth fibrous layer covered the silastic tube and the endplate of ATDS apparently (Figure 11). A small quantity of fluid flew out when the capsule was opened along the edge of the plate. The lumen of the T-shaped tube and pressure confined mechanism were
filled with aqueous humor and devoid of obstructions in all specimens examined, suggesting free flow of fluid.

DISCUSSION

ATDS was developed in accordance with preventing hypotony in routine glaucoma filtration surgery\textsuperscript{26}, and a new GDD must demonstrate consistent control over internal flowed. The pressure difference between the inlet and outlet of ATDS was mainly influenced by the aqueous flow rate, and produced by frictions of tube wall, resistance of sinuosity, small diameter and sudden dimension change.

According to Energy Conservation Law, conversion of one type of matter into another are always accompanied by the conversion of one form of energy into another. In this study, the potential energy (in form of pressure drop), tube resistance and kinetic energy of aqueous humor are conversed at any time if heat exchange is not under consideration. Consequently, if fluid flows at a steady rate, the pressure of inlet must be higher than that of the outlet to keep the potential energy converting to the kinetic energy. Pressure drop and flow rate influences each other, and there should be 3 conditions as follows when applying to ATDS: 1) as is shown in the hydrodynamic tests, an 8.5 mm Hg pressure drop will be produced by the pressure confined mechanism when aqueous humor is secreted and drained out at a stable flow rate of 2.5 \( \mu \)L/min, or IOP will be 8.5 mm Hg higher than the inner pressure of the filtration bleb. The success limit of 21 mm Hg needs a steady flow rate of 6.2 \( \mu \)L/min, which is higher than the physiological range of human aqueous secretion; 2) if the pressure difference between AC and filtration bleb is higher than 8.5 mm Hg, the flow rate will grow bigger than aqueous secretion, which will then cause pressure impairment to decrease the pressure drop until the balance of 8.5 mm Hg is back; 3) aqueous humor could flow outside under any pressure lower than 8.5 mm Hg but higher than its hydrostatic pressure. However, the lowest flow rate in outside under any pressure lower than 8.5 mm Hg but higher pressure impairment to decrease the pressure drop until the will grow bigger than aqueous secretion, which will then cause

The complications triggered by the surgery may have been attributable to the larger incision to AC, reject reactions and irritation of the T-shaped tube. The local evocation of iris root demonstrated the pressing from the tube. Although the AC angle in rabbit is longer and narrower than in human, the length and the outer diameter of the T-shaped tube should be decreased to reduce the contact and irritation, without hindering the blockage to the cornea incision. It is satisfactory that all the ATDSs were filled with aqueous humor without any obstructions. The micropores distributed to the horizontal tube are thought to be useful to increase drainage surface and avoid tube obstruction. Blockage may happen in some of the micropores, but others are still open to ensure the aqueous humor outflow.

A thin capsule was found around ATDS on day 60. The hygric granular inner surface with aqueous filled, relating to the hypoechoic space in the B-scan ultrasonography and the stable reduction of IOP, suggested a functional filtration bleb. Thin capsules were also found after glaucoma implant inserted in human or rabbit eyes, which consisted of lamellar collagen deposition surrounded by a granulomatous reaction with multinucleate giant cells\textsuperscript{28-29}. ATDS has an exclusive surgical procedure and it is not difficult to implant. This study has proved that ATDS could control IOP over internal flowed and provide consistent protection from hypotony in the early postoperative period. With this basal research, we are confident to improve the construction of ATDS and carry out further long-term animal studies including histopathological research in the future.

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Li-Jun Cui conceived and designed the study. Di-Chen Li designed and screened the pressure confined system, and made ATDS to be available. Jian Liu and Lei Zhang contributed to the Hydrodynamic tests. Li-Jun Cui, Lei Zhang and Yao Xing performed the experiments and wrote the paper. All authors read and approved the manuscript. Our team sincerely thanks to Mr. Jun-Tao Ning and Ms. Li-Hua Liu, School of Mechanical Engineering of Xi’an Jiaotong University, for assistance of mode building and manufacture of ATDS.

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