Toxocara optic neuropathy: clinical features and ocular findings

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Received: 2017-09-06 Accepted: 2017-11-21

Abstract

- We evaluated thirteen eyes of twelve patients diagnosed clinically and serologically with Toxocara optic neuropathy. Eleven patients had unilateral involvement and one patient had bilateral optic neuropathy. Eight patients (66.7%) had a possible infection source to Toxocara. Six patients (50%) had painless acute optic neuropathy. Ten eyes had asymmetric, sectorial optic disc edema with peripapillary infiltration and three eyes had diffuse optic disc edema. Eosinophilia was noted in five patients (41.7%) and optic nerve enhancement was observed in eight of eleven eyes (72.7%) with available orbit magnetic resonance imaging (MRI). Mean visual acuity significantly improved following treatment [mean logarithmic of the minimum angle of resolution (logMAR) 0.94±0.56 at baseline and 0.47±0.59 at the final (P=0.02)]. Asymmetric optic disc edema with a peripapillary lesion and a history of raw meat ingestion were important clues for diagnosing Toxocara optic neuropathy. Additionally, Toxocara IgG enzyme-linked immunosorbent assay (ELISA) test and evaluating eosinophil may be helpful for diagnosis.

- KEYWORDS: optic neuropathy; toxocariasis; ocular toxocariasis; Toxocara optic neuropathy

DOI:10.18240/ijo.2018.03.26

INTRODUCTION

Toxocariasis is one of the most common zoonotic infections worldwide[1-2]. The clinical presentation of toxocariasis in humans varies widely, with some patients having an asymptomatic infection and others developing severe organ damage. Symptom severity is dependent upon parasite load, larval migration site, and host inflammatory response. Moreover, systemic toxocariasis (also known as visceral larva migrans) and ocular toxocariasis can occur, which are particularly dependent upon on the involved organ[2-3]. Ocular toxocariasis is a clinically well-defined manifestation of an intraocular Toxocara larvae infection. This condition is an important cause of childhood vision loss, with ocular toxocariasis affecting both the retina and optic nerve. The ocular infection can also severely affect vision in adults, particularly in Asian adults because of cultural food habits[2,4]. Several case reports of toxocariasis involving the optic nerve have been published[5], but most ocular toxocariasis studies have focused on retinal damage, including retinal granuloma, epiretinal membrane, macular edema, and retinal detachment. Therefore, the clinical features, diagnosis, treatments, and disease courses of Toxocara optic neuropathy remain unclear. The current study evaluates distinguishing clinical features and the course of Toxocara optic neuropathy.

METHODS

All study conduct adhered to the tenets of the Declaration of Helsinki. This study was approved by the Institutional Review Board at Pusan National University Yangsan Hospital (Yangsan, Korea). We retrospectively evaluated Toxocara optic neuropathy between 2008 and 2016. Patients were diagnosed with Toxocara optic neuropathy if all of the following were present or true: 1) acute optic neuropathy [sudden onset of decreased visual acuity or visual field defect with a relative afferent pupillary defect (RAPD)]; 2) positive serum enzyme-linked immunosorbent assay (ELISA) titer for Toxocara canis IgG (ELISA titer of >0.250 considered serologically positive on a previous study showing 92.2% sensitivity and 86.6% specificity at an optical density =0.250)[6]; 3) all other possible...
causes of optic neuropathy ruled out, including ischemic optic neuropathy, retinal vessel occlusion, autoimmune disease, inflammatory conditions, cancer masquerade syndromes, and other infectious etiologies.

We investigated possible Toxocara infection sources and conducted a standardized interview, ensuring complete responses and participant comprehension to enhance interview validity. Standard interview questions included queries about ingestion of possible contaminated sources (raw animal liver, meat, and animal blood), history of pet ownership, and occupation-associated contact with animals or soil. Best-corrected visual acuity (BCVA), intraocular pressure (IOP), slit-lamp biomicroscopy findings, dilated fundus examination findings, Humphrey visual field testing results, anterior chamber/vitreous inflammatory status were reviewed in all subjects. Optic disc and fundus findings were photographed with a fundus camera (Kowa Co. Ltd., Tokyo, Japan) at each follow-up visit and changes in the optic disc and fundus were evaluated. Cross-sectional images of the optic disc and peripapillary area was obtained and evaluated using spectral-domain optical coherence tomography (SD-OCT; Spectralis, Heidelberg Engineering, Heidelberg, Germany). Subjects were also tested for eosinophilia (defined as >500 eosinophils/µL in peripheral blood or ≥5% of total white blood cell count) and underwent contrast-enhanced orbital magnetic resonance imaging (MRI).

Subjects with Toxocara optic neuropathy were treated with systemic corticosteroid therapy (intravenous steroid pulse therapy for 3d followed by tapering with oral prednisolone for 2wk), anti-parasitic therapy (400 mg albendazole twice a day for 2wk), or a combination of both treatments. Topical corticosteroid therapy (prednisolone acetate 1% four times a day) was also used when anterior chamber inflammation was present.

Treatment outcome was defined as the difference between initial and final visual acuity in patients who had at least 1-month follow-up. Snellen BCVA measurement was converted to the logarithmic of the minimum angle of resolution (logMAR) for all data analyses. Wilcoxon’s rank sum test was used to test statistical significance.

RESULTS AND DISCUSSION

Thirteen eyes of twelve patients were diagnosed with Toxocara optic neuropathy and demographic and clinical features were summarized in Table 1.

Ocular toxocariasis predominantly occurs in men\textsuperscript{[1,3,7-8]}.

However, our study showed that Toxocara optic neuropathy can also affect women in a similar manner. Most cases included in the current study were unilateral, but one subject did have bilateral involvement. Six subjects (50%) had ocular pain at the time of optic neuropathy presentation and the other six patients had painless acute optic neuropathy. We revealed a strong association between Toxocara optic neuropathy and ingestion of uncooked animal products. Eight patients (66.7%) had identification of a possible infection source, with two patients having a history of puppy/kitten exposure and six patients having history of ingesting raw animal liver, raw meat, or red ants. The remaining four patients (33.3%) had a nonspecific history. This information highlights the importance of asking adult optic neuropathy patients about their consumption of raw meat or liver. Unfortunately, we were not able to elucidate

### Table 1: Demographics and clinical findings of Toxocara optic neuropathy

<table>
<thead>
<tr>
<th>Patient No./age (y)/gender/eye</th>
<th>Infection source history</th>
<th>Ocular pain</th>
<th>Initial BCVA</th>
<th>Final BCVA</th>
<th>Asymmetric optic disc edema/ peripapillary lesion</th>
<th>Posterior pole</th>
<th>Visual field defect</th>
<th>Eosinophilia</th>
<th>MRI enhance</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>1/36/M/L</td>
<td>+</td>
<td>CF</td>
<td>CF</td>
<td>+/+</td>
<td>Chorioretinitis, HE</td>
<td>Diffuse</td>
<td>+</td>
<td>-</td>
<td>A</td>
<td></td>
</tr>
<tr>
<td>2/49/M/R</td>
<td>+</td>
<td>20/100, HM</td>
<td>20/30,20/30</td>
<td>+/+, +/-</td>
<td>-</td>
<td>NA</td>
<td>-</td>
<td>+/+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>3/49/F/R</td>
<td>+</td>
<td>NLP</td>
<td>NLP</td>
<td>+/+</td>
<td>-</td>
<td>Diffuse</td>
<td>-</td>
<td>+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>4/71/F/R</td>
<td>-</td>
<td>20/320</td>
<td>NA</td>
<td>+/-</td>
<td>Vascularis, HE</td>
<td>Inferior</td>
<td>+</td>
<td>-</td>
<td>S</td>
<td></td>
</tr>
<tr>
<td>5/71/M/R</td>
<td>-</td>
<td>20/25</td>
<td>NA</td>
<td>+/-</td>
<td>-</td>
<td>Superior</td>
<td>-</td>
<td>-</td>
<td>S</td>
<td></td>
</tr>
<tr>
<td>6/60/M/R</td>
<td>+</td>
<td>20/100</td>
<td>20/32</td>
<td>+/-</td>
<td>ME, HE</td>
<td>Superior</td>
<td>+</td>
<td>+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>7/52/M/R</td>
<td>+</td>
<td>20/200</td>
<td>20/30</td>
<td>+/-</td>
<td>ME, HE</td>
<td>Diffuse</td>
<td>-</td>
<td>+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>8/31/F/R</td>
<td>-</td>
<td>20/2000</td>
<td>20/125</td>
<td>+/-</td>
<td>Vascularis, ME, HE</td>
<td>Diffuse</td>
<td>+</td>
<td>+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>9/60/F/L</td>
<td>-</td>
<td>20/25</td>
<td>20/25</td>
<td>+/-</td>
<td>-</td>
<td>Diffuse</td>
<td>+</td>
<td>+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>10/60/F/L</td>
<td>+</td>
<td>20/32</td>
<td>20/40</td>
<td>+/-</td>
<td>-</td>
<td>Quadranopia</td>
<td>-</td>
<td>+</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>11/47/M/L</td>
<td>+</td>
<td>20/100</td>
<td>20/20</td>
<td>+/-</td>
<td>ME, HE</td>
<td>Diffuse</td>
<td>-</td>
<td>NA</td>
<td>S+A</td>
<td></td>
</tr>
<tr>
<td>12/60/M/R</td>
<td>+</td>
<td>20/160</td>
<td>20/32</td>
<td>+/-</td>
<td>-</td>
<td>Diffuse</td>
<td>-</td>
<td>NA</td>
<td>S+A</td>
<td></td>
</tr>
</tbody>
</table>

CF: Counting fingers; HM: Hand movements; NLP: No light perceptions; ME: Macular edema; HE: Hard exudation; NA: Non-applicable; S: Corticosteroid pulse treatment with slow tapering; A: Albedazole. Eosinophil counts >5% (normal range, 1% to 5%).
the timing between raw meat or liver ingestion and ocular symptom onset. Previous studies have shown that *Toxocara* larvae can survive in human tissues for over 10 years, therefore, when *Toxocara* optic neuropathy is suspected, the patient should be asked about raw meat ingestion up to 10 years ago[5,9]. However, a definitive diagnosis of ocular toxocariasis can be obtained via histological confirmation of the *Toxocara* larva or its fragments in infected tissue samples. Collection of intraocular tissue is difficult and rarely warranted in clinical practice. Thus, *Toxocara* IgG ELISA testing is likely an important diagnostic tool and should be done immediately in patients with suspected *Toxocara* optic neuropathy. Ahn et al[2] reported that eosinophil was not as helpful as serum anti-*Toxocara* IgG test in ocular toxocariasis diagnosis (only 11.6% had eosinophilia). However, eosinophilia may indicate the presence of *Toxocara* larvae, as shown in previous reports[1,7], and our study showed 41.6% patient had eosinophilia. Therefore, we recommended that eosinophilia would be additional diagnostic clues for *Toxocara* optic neuropathy diagnosis if present.

All eyes had optic disc edema; ten eyes had asymmetric and/or sectorial optic disc edema with associated subretinal fluid or peripapillary infiltration (Figure 1A). SD-OCT image of asymmetric optic disc edema with peripapillary lesions showed optic disc edema with subretinal fluid and moderately hyper-reflective mass like lesion, which had posterior shadowing (Figure 1B). The remaining three eyes had diffuse, generalized optic disc edema without peripapillary lesion. Six eyes had retinal lesions with optic disc swelling because of chorioretinitis, retinal vasculitis, macular edema, and hard exudates (Figure 1C). Yang et al[5] published a *Toxocara* optic neuropathy case series and showed that optic disc swelling was present in all patients, but ranged from subtle to severe. Circumpapillary lesions were presented in all patients and eventually developed retinal lesions[5]. These clinical signs were also present in our *Toxocara* optic neuropathy patients, we therefore highlight that asymmetric and/or sectorial optic disc edema with peripapillary lesion may be important clinical clue in *Toxocara* optic neuropathy. Interestingly, the current study showed that *Toxocara* optic neuropathy can present as a diffuse optic disc swelling without peripapillary and retinal lesions, in addition, some patients had no significant MRI enhancement. These differences in presentation may represent differences in disease severity and/or stage. Disease severity depends not only on the number of larvae ingested, but also on the severity of a patient’s allergic reaction[10-11]. As a result, patients with atopy may experience more severe toxocariasis[10-11]. Pathologic manifestations result from the immune-mediated inflammatory response directed against the excretory-secretory larval antigens. These antigens are released from larvae’s outer epicuticle coat, which readily sloughs off when bound by specific antibodies[12]. These antigens are a mix of glycoproteins, including a potent allergenic component named tubulin alpha chain -1 (TBA-1)[11,13]. Hayashi et al[13] showed that the optic nerve may be a migratory route of *Toxocara canis* from the brain to the eye in an animal model. Therefore, the optic disc may appear normal while *Toxocara canis* migrate within the nerve, before development of optic disc edema. As a result, patients suspected of having *Toxocara* that affects the optic nerve should be closely monitored.

Optic nerve enhancement was observed on orbital MRI in eight of eleven eyes (72.7%) with available MRI. Optic nerve enhancement was localized to the orbital portion of optic nerve in all patients. There were no abnormal findings in the extra-orbit portion of optic nerve and brain parenchyma of any patient. Humphrey visual field testing had been performed in ten patients. Testing results most commonly showed a diffuse...
visual field defect in eyes with *Toxocara* optic neuropathy (Table 1).
Nine patients were treated with intravenous pulse corticosteroid therapy and albendazole combination therapy, two patients were treated with intravenous pulse corticosteroid therapy, and one patient was treated with albendazole mono therapy. Eleven eyes of ten patients were included in the comparison between initial and final visual acuity. Visual acuity significantly improved following treatment, mean BCVA was 0.94±0.56 logMAR [range: no light perception (NLP) to 20/20] at baseline and 0.47±0.59 (range: NLP to 20/20) at the final visit \( (P=0.02) \). Four eyes (36.4%) had severe vision loss (worse then 20/200) at the initial visit, and then two eyes among four eyes with severe vision loss at initial visit remained severe vision loss at the final visit. Treatment of toxocariasis has been standardized with steroid treatment protocols, but efficacy of anti-helminthics has not been established\(^{[16]}\). Nevertheless, we could not compare treatment outcomes after different treatment regimen, Ahn et al\(^{[3]}\) reported that albendazole and oral prednisolone combination therapy significantly lowered the 6-month recurrence rate compared with corticosteroid monotherapy. We therefore recommend combined corticosteroid and albendazole treatment may effective for reducing optic nerve inflammation and recurrence.

Our study had several limitations. First, subject interviews were based on standardized questionnaires administered at the time of initial patient presentation. Using careful documentation, we had to rely on patient interviews to identify probable *Toxocara* infection sources. Second, treatment outcomes were not examined by treatment method and it is possible that visual outcomes may differ for each treatment type (corticosteroid, anti-helminthics, and combination therapy). Further prospective, randomized trials are needed to compare treatment outcomes among the various treatment protocols. In conclusion, asymmetric optic disc swelling in the presence of a peripapillary and/or retinal lesion, and a history of raw meat ingestion were important clinical findings in adult patient with *Toxocara* optic neuropathy. Corticosteroid and albendazole combination therapy would help to prevent serious visual impairment in patients with *Toxocara* optic neuropathy.

**ACKNOWLEDGEMENTS**

**Conflicts of Interest:** Choi KD, None; Choi JH, None; Choi SY, None; Jung JH, None.

**REFERENCES**