Neurofibromatosis type 2 gene mutation and progesterone receptor messenger RNA expression in the pathogenesis of sporadic orbitocranial meningioma

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Abstract

- **AIM:** To investigate neurofibromatosis type 2 (NF2) gene mutation at mRNA levels in sporadic orbitocranial meningioma and its association with progesterone receptor (PR) mRNA expression.
- **METHODS:** This was a case-control study. Thirty-four sporadic meningioma patients with no familial NF2-related meningioma history were recruited. They were interviewed for their obstetric, gynecologic, and contraception history. PR investigation was performed with real-time polymerase chain reaction (PCR). NF2 mutation was investigated using Qbiomarker Somatic Mutation PCR Assay at NF2 mRNA level after its cDNA extraction (four mRNA mutation cytoband coordinates for nucleotide change: c.634C>T/p.Q212, c.655G>A/p.V219M, c.784C>T/p.R262 and c.1228C>T/p. Q410). A
- **RESULTS:** After mutation analysis at mRNA level, NF2 gene mutation was found in 35.29% patients. Non-mutation group was strongly associated with exogenous hormonal exposure (non-mutation vs mutation: 95.5% vs 83.3%, P<0.001). PR mRNA was found significantly lower in non-mutation group (P=0.033) which presumed as long term exogenous progesterone exposure. However, mutation group was associated with higher rate of progression to grade II (mutation vs non-mutation, 18.2% vs 5%, P<0.001) and was associated more in fibrous and anaplastic tumor tissue.
- **CONCLUSION:** NF2 mutation-meningioma is associated with higher grade of meningioma. Non NF2 mutation-meningioma is strongly associated with exogenous progesterone exposure and lower PR expression.
- **KEYWORDS:** orbitocranial meningioma; neurofibromatosis type 2; progesterone receptor; hormonal contraception; real time PCR

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INTRODUCTION

Meningioma is a primary brain tumor derived from arachnoid cells[1-3]. Meningiomas have slow growth properties that make them undetectable in causing symptoms of tumor compression[1]. Orbitocranial meningiomas can be classified by their location into primary, inferorbital and spheno-orbital optic meningiomas[4]. Rapid growth, infiltration to adjacent structures (connective tissue, bone, and brain tissue) can also occur in benign to malignant progression[2]. Progression of meningiomas has been known as the result of NF2 gene mutation[5]. NF2 gene analysis study has found that somatic mutation of meningioma (monosomy and chromosome deletion 22q) underlies and supports the hypothesis that NF2 serves as a tumor suppressor in meningioma tumorigenesis[6-7].
Chromosomal instability in meningioma can be used as a marker for benign to malignant transformation\textsuperscript{(9)}. NF2 gene mutation on chromosome 22 leads to phenotypic changes from arachnoid cell hyperplasia to benign meningiomas with 70\%-80\% fibroblastic (and transitional) histopathologic types and only 25\% meningothelial type\textsuperscript{(9)}. Possible role of steroid hormones and their receptors in the pathogenesis of meningiomas is remain unclear and reveals inconclusive conclusions regarding its developmental mechanisms and role in the risk of recurrent cases\textsuperscript{(10-12)}. The progesterone receptor (PR) status has been known to be associated with a gene mutation on chromosome 22 (at a locus adjacent to the NF2 gene) which has been identified as the initial sign of meningioma tumorigenesis\textsuperscript{(13)}. Those findings suggest that PR status could be used as clinical marker of genetic mutation of meningiomas which requires further study. Therefore, this study aimed to investigate the NF2 gene mutation at mRNA level and reveal the association between NF2 gene mutation and progesterone and estrogen receptors (ER) mRNA expression in meningioma. It might reveal whether orbitalcranial meningioma is associated by intrinsic sporadic NF2 mutation or due to extrinsic factors (hormonal contraceptives exposure) as resembled by hormone receptors status (PR and ER).

**SUBJECTS AND METHODS**

**Ethical Approval** This study followed the tenets of the Declaration of Helsinki (2008) and was approved by Ethical Review Board of the Faculty of Medicine, Public Health and Nursing, Universitas Gadjah Mada (Approval No: KE/FK/178/EC/2016). Written informed consent was obtained from all study participants.

**Research Design and Subjects** This study was an analytic observational study with case-control design. The tissue samples were taken consecutively from histopathologic specimen of meningioma tissue originating from patients diagnosed with sporadic orbitalcranial meningioma (not associated with any familial neurofibromatosis 2) underwent lateral orbitotomy at Dr. Sardjito General Hospital and dr. Yap Eye Hospital, Yogyakarta, Indonesia from 2010 to 2016. Patients’ basic characteristics were taken from the medical records which were corresponding to their histopathologic specimens. Patients whose tissues were included in the inclusion criteria were then contacted for interviews for their history of obstetric and gynecologic as well as their history of hormonal contraception using a questionnaire. Exclusion criteria of this study were patients who did not meet inclusion criteria, damaged paraffin blocks, insufficient cutting, incomplete medical record data and the patient cannot be contacted for interviews.

**Quantitative Polymerase Chain Reaction** Paraffin blocks of meningioma tissue were cut as thick as 4 \(\mu\)m. NF2 mutation was examined using quantitative PCR with Qbiomarker Somatic Mutation PCR Assay (4 cytoband coordinates: c.634C>T/p.Q212, c.655G>A/p.V219M, c.784C>T/p.R262, and c.1228C>T/p.Q410). Quantitative PCR examination was also performed to determine ER and PR expression after RNA extraction, amplification of cDNA was then performed for cDNA quantification (DThite4, DNA-Technolog, Russia). Primer sets were GAPDH as a reference (GAPDH forward: 5'-GCA TCC TGG GCT ACA CTG AG-3'; GAPDH reverse: 5'-TCC ACC ACC CTG TTG CTG TA-3'), PR (forward: 5'-AGC TCA TGA AGG CAA TTG GTT T-3'; reverse: 5'-ACA AGA TCA TGC AAG TTA TCA AGA AGT T-3'), ER\(\alpha\) (forward: 5'-TCA CAT CTG TAT GCG GAA CC-3'; reverse: 5'-CGT AAC ACT TCC GAA GTC GG-3'), ER\(\beta\) (forward: 5'-AGA CAT GAG AGC TGC CAA CC-3'; reverse: 5'-GCC AGGCAC ATT CTA GAA GG-3') and NF2 (forward: 5'-CCC CCA ACT CCT CTT GCC CCC CTG-3')

**Statistical Analysis** Continuous variables (e.g. age, gynecological history, and history of hormonal contraceptive use as well as PR and ER expression) were tested by Mann-Whitney test and categorical variables (sex, marital status, level of education, occupation, income, degree and histopathologic type of tumor) were tested using Fisher-Exact test. Odds ratio (OR) was obtained using univariate and multivariate logistic regression test. The \(P<0.05\) was considered significant.

**RESULTS** Thirty-four orbitalcranial meningiomas tissues met the inclusion criteria. NF2 gene mutations were present in 35.29\% (12 cases). The mean age of all subjects was 45.08±6.51y. There were no significant differences in age, education, occupation, family history of cancer, and eye complaints between the 2 groups (Table 1). The mean age of menarche of all subjects was 13.56±1.44y. There was no significant difference in age of menarche, menstrual cycle, menstrual period, and parity between 2 groups. There was a statistically significant difference in the proportion of irregular menstruation between 2 groups (\(P<0.001\)) as well as in the proportion of contraceptive acceptors between mutation and non-mutation groups [non-acceptor in the mutation group (16.7\%) vs non-mutation (4.5\%), \(P=0.001\); Table 2]. There was statistically significant difference in the proportion of contraceptive use
Table 1 Subject characteristics

<table>
<thead>
<tr>
<th>Variables</th>
<th>Mutation</th>
<th>Non-mutation</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (y), mean±SD</td>
<td>47.17±6.44</td>
<td>43.95±6.40</td>
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<tr>
<td>Elementary</td>
<td>5 (41.7)</td>
<td>7 (31.8)</td>
<td></td>
</tr>
<tr>
<td>Junior</td>
<td>2 (16.7)</td>
<td>5 (22.7)</td>
<td></td>
</tr>
<tr>
<td>Junior high</td>
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<td>7 (31.8)</td>
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</tr>
<tr>
<td>Diploma-3</td>
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<td>1 (4.5)</td>
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<td>2 (9.1)</td>
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<tr>
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<tr>
<td>None</td>
<td>9 (75)</td>
<td>11 (50)</td>
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<tr>
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<td>4 (18.2)</td>
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<td>Monthly income</td>
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<td>&lt;$ 100</td>
<td>7 (53.3)</td>
<td>7 (31.8)</td>
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<td>$100-500</td>
<td>5 (41.7)</td>
<td>12 (54.5)</td>
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<tr>
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<td>Complaints</td>
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<td>Proptosis</td>
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<td>Blurred vision</td>
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<td>2 (9.1)</td>
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<td>Headache</td>
<td>6 (50.0)</td>
<td>8 (36.4)</td>
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<td>Others</td>
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<tr>
<td>No</td>
<td>12 (100)</td>
<td>16 (72.7)</td>
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<td>History of breast cancer</td>
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<td>1 (4.5)</td>
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<tr>
<td>No</td>
<td>11 (91.7)</td>
<td>21 (95.5)</td>
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Table 2 Obstetric-gynecologic and hormonal contraception history

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<th>Variables</th>
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<th>Non-mutation</th>
<th>P</th>
</tr>
</thead>
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<tr>
<td>Obstetric-gynecologic history</td>
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<td></td>
<td></td>
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<tr>
<td>Age of menarche (y), mean±SD</td>
<td>13.92±1.78</td>
<td>13.36±1.22</td>
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<td>Menstrual cycle</td>
<td>0.399</td>
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<td>28d</td>
<td>11 (91.7)</td>
<td>17 (77.3)</td>
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<td>&gt;28d</td>
<td>0</td>
<td>3 (13.6)</td>
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<td>Irregular</td>
<td>1 (8.3)</td>
<td>2 (9.1)</td>
<td></td>
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<td>Menstruation</td>
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<td></td>
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<td>6 (50)</td>
<td>14 (63.6)</td>
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<td>8 (36.4)</td>
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<td>Irregular menstruation</td>
<td>&lt;0.001*</td>
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<td>8 (80)</td>
<td>19 (90.5)</td>
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<td>No</td>
<td>2 (20)</td>
<td>2 (9.5)</td>
<td></td>
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<tr>
<td>Months of irregularity, mean±SD</td>
<td>187.5±124.03</td>
<td>139.06±112.29</td>
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<td>Parity, mean±SD</td>
<td>2.33±1.15</td>
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<td>0.071</td>
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<td>Hormonal contraception history</td>
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<tr>
<td>Acceptor</td>
<td>&lt;0.001*</td>
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<tr>
<td>Yes</td>
<td>10 (83.3)</td>
<td>21 (95.5)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>2 (16.7)</td>
<td>1 (4.5)</td>
<td></td>
</tr>
<tr>
<td>Onset (y), mean±SD</td>
<td>25.8±4.8</td>
<td>25.00±5.1</td>
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<tr>
<td>Types</td>
<td>&lt;0.001*</td>
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<tr>
<td>3 monthly injection</td>
<td>10 (100)</td>
<td>19 (90.5)</td>
<td></td>
</tr>
<tr>
<td>Oral</td>
<td>0</td>
<td>2 (9.5)</td>
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</tr>
<tr>
<td>Duration (mo), mean±SD</td>
<td>163.8±99.76</td>
<td>149.0±63.7</td>
<td>0.849</td>
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</tbody>
</table>

Significant statistically.

Table 3 Degree and type of histopathology of tumor

<table>
<thead>
<tr>
<th>Variables</th>
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<th>Non-mutation</th>
<th>P</th>
</tr>
</thead>
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<tr>
<td>Degree</td>
<td>&lt;0.001*</td>
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<td>WHO grade I</td>
<td>9 (81.8)</td>
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<td>WHO grade II</td>
<td>2 (18.2)</td>
<td>1 (5)</td>
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<tr>
<td>Histopathology</td>
<td></td>
<td></td>
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<tr>
<td>Meningothelial</td>
<td>3 (27.3)</td>
<td>7 (35.0)</td>
<td>0.804</td>
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<tr>
<td>Microcystic</td>
<td>1 (9.1)</td>
<td>2 (10.0)</td>
<td>0.022*</td>
</tr>
<tr>
<td>Fibrous</td>
<td>2 (18.2)</td>
<td>2 (10.0)</td>
<td>0.039*</td>
</tr>
<tr>
<td>Transitional</td>
<td>3 (27.3)</td>
<td>6 (30.0)</td>
<td>0.607</td>
</tr>
<tr>
<td>Psammomatous</td>
<td>0</td>
<td>1 (5)</td>
<td>0.003*</td>
</tr>
<tr>
<td>Angiomatous</td>
<td>0</td>
<td>1 (5)</td>
<td>0.003*</td>
</tr>
<tr>
<td>Anaplastic</td>
<td>2 (18.2)</td>
<td>1 (5)</td>
<td>0.006*</td>
</tr>
</tbody>
</table>

Significant statistically.

The present study was limited to detecting the presence or absence of NF2 mutations (qualitatively) from the 4 cytoband coordinates for nucleotide change (c.634C>T/p.Q212, c.655G>A/p.V219M, c.784C>T/p.R262, and c.1228C>T/p.Q410). NF2 mutations were found in c.634C>T/p.Q212 (50%) and c.1228C>T/p.Q410 (41.67%). NF2 expression was 30.33±1.19 and was not associated with NF2 mutation (P=0.294; OR=0.697, 95%CI: 0.355-1.368). NF2 mutations in cytoband c.655G>A/p.V219M and c.784C>T/p.R262 were significantly associated with meningothelial and transitional histopathology type (Table 5). In this study, there were no significant confounding factors statistically interfering with the results of the study (Table 6).
DISCUSSION
This study was the first study investigating NF2 mutation at the level of mRNA and its association with mRNA PR which might become an important method for future investigation using serum (“liquid biopsy”). The result of NF2 mutation in the present study was in line with a previous study in which the mutation of NF2 gene (on chromosome 22) was found in 30% to 70% of sporadic meningiomas[8]. The most common genetic alteration found in meningiomas involving NF2 gene on the 22q chromosome where the incidence of NF2 mutations can in the form of insertion, deletion, or nonsense mutation[14].
The loss of the NF2 gene product (merlin) is considered to be an early event in the pathogenesis of sporadic meningiomas. Tumor suppressor gene inactivation might need bi-allelic NF2 inactivation that is NF2 intrinsic mutation and exogenous events[14] such as chronic exposure to exogenous progesterone.
The orbitocranial meningioma is arising from the skull base, so called sphenoid-orbital meningiomas, which is more associated with the incidence of chromosome 22q mutations and has more aggressive clinical course than meningiomas on the cerebrum, cerebellum and lateral skull surfaces[4]. However, intracranial meningiomas might exhibit more heterogeneous and complex chromosome abnormalities associated with more than one tumor cell clone[15].

**Table 4** ER serum and tissue expression

<table>
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<tr>
<th>Variables</th>
<th>Mutation</th>
<th>Non-mutation</th>
<th>P</th>
<th>OR (95%CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ERα</td>
<td>Serum</td>
<td>29.83±0.83</td>
<td>30.95±1.70</td>
<td>0.079</td>
</tr>
<tr>
<td>Tissue</td>
<td>34.14±0.74</td>
<td>34.68±0.57</td>
<td>0.060</td>
<td>4.261 (1,023-17.74)</td>
</tr>
<tr>
<td>ERβ</td>
<td>Serum</td>
<td>23.61±2.54</td>
<td>22.78±2.21</td>
<td>0.438</td>
</tr>
<tr>
<td>Tissue</td>
<td>33.93±1.23</td>
<td>34.10±1.05</td>
<td>0.613</td>
<td>1.149 (0.604-2.188)</td>
</tr>
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</table>

**Table 5** Histopathology of meningioma and cytoband coordinates for nucleotide change of NF2 mutation

<table>
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<th></th>
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<tbody>
<tr>
<td>Meningothelial</td>
<td>0.388</td>
<td>0.012</td>
<td>0.039</td>
<td>0.267</td>
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<tr>
<td>Microcystic</td>
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<td>0.625</td>
<td>1.000</td>
<td>0.727</td>
</tr>
<tr>
<td>Fibrous</td>
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<td>0.375</td>
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<td>Transitional</td>
<td>0.549</td>
<td>0.008</td>
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<tr>
<td>Psammomatous</td>
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<td>1.000</td>
<td>1.000</td>
<td>0.219</td>
</tr>
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<td>Angiomatous</td>
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<td>1.000</td>
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<tr>
<td>Anaplastic</td>
<td>0.219</td>
<td>1.000</td>
<td>1.000</td>
<td>0.453</td>
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*Statistically significant.

**Table 6** Confounding analysis

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<th>Multivariate analysis</th>
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<tr>
<td></td>
<td>P</td>
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<td>Onset of contraception use</td>
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<td>Type of hormonal contraception</td>
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<td>Age of menarche</td>
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<tr>
<td>Menstrual cycle</td>
<td>0.497</td>
<td>1.585</td>
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</table>

**Figure 1** PR mRNA expression in NF2 mutation and non NF2 mutation meningioma  Significant mean difference of serum NF2 between mutation and non-mutation groups (P=0.033).
Gynecological history is a marker of endogenous hormonal exposure and the use of hormonal contraceptives is a marker of exogenous hormonal exposure. However, in this study it was found that endogenous hormonal factors (age of menarche, menstrual cycle and menstrual period, and parity) were not related to NF2 mutation. A study by Claus et al.[12] revealed that age of menarche is not associated with the risk of meningioma. The evidence related to the role of endogenous progesterone in the meningioma has been revealed by the finding of association of PR+ cases with the incidence of meningioma[16]. There is a sign of the role of endogenous progesterone hormone that is the worsening symptoms during the luteal phase of the menstrual cycle and during pregnancy. However, previous research had shown the associated risk of meningioma in patients with hormonal contraceptive use (oral, sub-dermal implants, injection, and intrauterine devices with hormones)[17] as well as in users of hormone replacement therapy.

The role of sex hormones was revealed by the finding that 88% of primary meningiomas are positive PR and 40% positive ER[18]. The hormone receptor association with NF2 gene mutation in the tumorigenesis of meningioma was known from the presence of a chromosome 22q mutation with the condition of ER-/PR- or ER+/PR+[13]. Negative or low PR expression might be associated with a history of chronic exposure to exogenous progesterone. Therefore, meningioma tumorigenesis in the non-mutation group might be assumed to be due to a chronic use of exogenous progesterone (hormonal contraceptives). While in the tumorigenesis in mutation group might be underlined merely on intrinsic factor (intrinsic NF2 gene mutation). The loss of PR is associated with a higher degree of meningioma which can be identified by MIB-1/Ki-67[19]. Increased MIB-1 was also associated with an increased recurrence risk in individuals who had previously been performed tumor resection[19].

NF2 mutations has been known to be associated with a higher degree of meningioma and increased risk of recurrence rate[20]. In the previous study, NF2 mutation was found in 17% to 38% grade II meningiomas and characterized by mutations not only on the 22q chromosome but also at 1p, 6q, and 14q[8]. Atypical meningiomas are associated with loss of 1p, 6q, 10q, 14q, 17p and 18q and gains at 20q, 12q, 15q, 1q, 9q and 17q[16]. The most common types of histopathologic (subtypes) in meningiomas are fibrous, transitional, and meningothelial. NF2 mutation was associated with fibroblastic and transitional histopathologic types[20]. In this study the highest proportion of meningothelial and transitional is 27.3%, followed by fibrous and anaplastic as much as 18.2%. NF2 mutations are known to be a result in increased production of collagen and extracellular matrix causing a histopathologic type associated with collagen-containing meningiomas such as fibrous and transitional[13].

In the present study the mutation characteristics such as homozygous deletions or neutral copies of LOH (loss of heterozygosity) were not investigated. The loss of allele loss on chromosome 22q12.2 (which includes the NF2 gene) is found in 40%-70% of sporadic meningiomas and most of the NF2-associated meningiomas[19]. In a previous study, it was found that only 15% meningiomas without LOH chromosome 22 had NF2 mutations[21]. Meningioma pathogenesis was not only independent of NF2 mutation, but also strongly associated with exogenous progesterone exposure. Benign to malignant meningioma progression was found to be due to NF2 mutation. Mutation analysis at mRNA level was a simple and reliable method to investigate mutation using patients’ serum and promising method for so called a “liquid biopsy”. NF2 mutations can be used for early identification of tumor progression, planning of surgical intervention to prevent the development of meningiomas thereby reducing morbidity and mortality.

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