

Distribution of pathogenic bacteria and antimicrobial sensitivity of eye infections in Suzhou

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Abstract

• **AIM:** To investigate the types of bacteria in patients with eye infections in Suzhou and their drug resistance to commonly used antibacterial drugs.

• **METHODS:** The clinical data of 155 patients were retrospectively collected in this study, and the pathogenic bacteria species and drug resistance of each pathogenic bacteria were analyzed.

• **RESULTS:** Among the 155 patients (age from 12 to 87 years old, with an average age of 57, 99 males and 56 females) with eye infections (160 eyes: 74 in the left eye, 76 in the right eye and 5 in both eyes, all of which were exogenous), 71 (45.81%) strains were gram-positive bacteria, 23 (14.84%) strains were gram-negative bacteria and 61 (39.35%) strains were fungi. Gram-positive bacteria were highly resistant to penicillin and erythromycin (78.87% and 46.48% respectively), but least resistant to vancomycin at 0. Gram-negative bacteria were highly resistant to

cefotaxime and compound sulfamethoxazole (100% and 95.65% respectively), but least resistant to meropenem at 0. Comparison of the resistance of gram-positive and gram-negative bacteria to some drugs revealed statistically significant differences ($P < 0.05$) in the resistance of both to cefotaxime, cotrimoxazole, levofloxacin, cefuroxime, ceftriaxone and ceftazidime, and both had higher rates of resistance to gram-negative bacteria than to gram-positive bacteria. The distribution of bacterial infection strains showed that *Staphylococcus epidermidis* was the most common strain in the conjunctiva, cornea, aqueous humor or vitreous body and other eye parts. Besides, *Fusarium* and *Pseudomonas aeruginosa* were also among the most common strains of conjunctival and corneal infections.

• **CONCLUSION:** Gram-positive bacteria are the dominant bacteria in eye infections, followed by gram-negative bacteria and fungi. Considering the resistance of gram-negative bacteria to multiple drugs, monitoring of bacteria should be strengthened in eye bacterial infections for effective prevention and control to reduce complications caused by eye infections.

• **KEYWORDS:** eye infection; pathogenic bacteria; drug resistance; antimicrobial sensitivity test

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INTRODUCTION

Eye infections refer to common and frequently-occurring ophthalmic diseases caused by various bacterial infections, which are generally caused by pathogenic factors such as bacteria, fungi and viruses, of which bacterial infections are the most common^[1-2]. Eye infections have a close bearing on various risk factors, such as trauma, surgery, contact lens wearing, age, dry eye, chronic nasolacrimal duct obstruction, and previous eye infections^[3-4], with varying degrees of prognosis, and in severe cases, can be vision-threatening^[5]. With keratitis and conjunctivitis as the two most

common types of infections^[6-7], eye infections can damage patients' eye tissues, resulting in decreased vision and, in severe cases, corneal perforation and eyeball atrophy^[8], which can seriously affect patients' normal life and even lead to blindness^[9-10].

Despite the natural resistance of human corneas to infection, factors such as trauma, corneal surgery, contact lens wearing^[11], ocular surface diseases^[12], systemic diseases^[13], and immunosuppression can alter the defense mechanisms of the ocular surface, resulting in bacterial invasion of the cornea. If left undiagnosed and untreated, keratitis may develop into endophthalmitis^[14], a serious eye infection caused by bacteria and other microorganisms entering the vitreous body. In a study by Wan *et al*^[15], the etiological characteristics of 531 cases of suppurative endophthalmitis in Qingdao Eye Hospital of Shandong Eye Institute from January 2006 to December 2015 were retrospectively analyzed. A total of 224 strains of pathogenic bacteria were obtained in culture, of which gram-positive bacteria (54%) were predominant, with *Staphylococcus epidermidis* (25%) being the most common, followed by fungi (29.5%) and gram-negative bacteria (16.5%). Bacterial conjunctivitis, which accounts for about 50%-70% of infectious conjunctivitis^[16], affects not only the conjunctiva but also adjacent tissue structures and is a potential risk factor for other intraocular or extraocular infections^[17]. About 90% of individuals can isolate bacteria from conjunctival sac, including *Staphylococcus epidermidis*, *Diphtheroid bacillus* and *Propionibacterium acnes*, which deter other pathogenic bacteria from attacking by releasing antibiotic-like substances and metabolites. Infection occurs in a situation where the defenses of the host are weaker than the invasion ability of pathogenic bacteria^[18]. Yang *et al*^[19] reported *Streptococcus pneumoniae*, *Haemophilus influenzae* and *Staphylococcus aureus* as common causative agents of acute bacterial conjunctivitis, whose incubation period and duration of infection depends on the organismal status of the infected person, with infection latency and duration depending on the organism status of the infected person.

Socioeconomic development and environmental changes have witnessed an upward trend in the incidence of eye infections^[20-22]. Eye infections are generally treated with antibacterial drugs, but their types and biological characteristics are constantly changing with the widespread use of antibacterial drugs and the popularity of contact lenses. Accordingly, the therapeutic effect of antibacterial drugs on bacterial infections and bacterial resistance are constantly changing as well^[23]. Given the increased difficulty in the treatment and diagnosis of bacterial eye infection, treatment with effective antimicrobial drugs at an early stage of eye infection is of great importance for the preservation and restoration of visual function in patients^[24-25].

For a better treatment of patients with ocular bacterial infections, a periodic summary evaluation of eye pathogenic bacterial resistance is needed to provide a scientific basis for the diagnosis of eye infections and the use of antimicrobial drugs. In this study, 155 patients admitted to Lixiang Eye Hospital of Soochow University from January 2020 to March 2023 were retrospectively collected, and their clinical data were retrospectively analyzed to figure out the types, composition and drug resistance of bacterial infections, so as to provide a basis for the rational use of antibacterial drugs in clinic.

SUBJECTS AND METHODS

Ethical Approval This study was conducted in accordance with the Declaration of Helsinki and approved by the Research Ethics Committee of Lixiang Eye Hospital of Soochow University (ethical batch number: SLER2023101), and informed consent was obtained from all participants and their guardians. All methods were carried out in accordance with relevant guidelines and regulations.

Subjects The clinical data of 155 patients admitted to Lixiang Eye Hospital of Soochow University in Suzhou from January 2020 to March 2023 were retrospectively collected, including corneal specimens (108 cases), lacrimal sac specimens (20 cases), aqueous humor or vitreous body (17 cases) and other parts (10 cases). Inclusion criteria: 1) clinically diagnosed diseases such as keratitis, eyelid/orbital cellulitis, endophthalmitis, lacrimal gland and lacrimal duct infection; 2) clinical microbial culture specimens of conjunctival sac secretions, lacrimal passage specimens, conjunctival/corneal scrapings, aqueous humor, vitreous body, foreign bodies caused by ocular trauma and ocular pathological tissues. Exclusion criteria: 1) non-ocular infectious diseases; 2) microbial culture specimens that are non-ocular biological specimens, such as urine and sputum; 3) duplicate strains isolated from the same part of the same case and suspected contaminated strains. Specimens were collected in strict accordance with the aseptic operating procedures and sent to the microbiology laboratory of the Department of Laboratory Medicine for isolation and identification.

Specimen Collection Conjunctival sac specimens were collected as follows: the patient was placed in a sitting position and instructed to gaze upward and turn the lower eyelid. A saline-soaked cotton swab was then used to gently wipe the conjunctival sac of the lower lid, including the medial canthus. After sampling, bacterial culture was performed. Corneal specimens were collected as follows: the patient was placed in the lying position, and after surface anesthesia, necrotic tissue was first scraped from the surface of the corneal ulcer under the microscope to expose the corneal lesion. Then, as much of the corneal tissue as possible was scraped with an ophthalmic scalpel and cultured with bacteria. Eyelid margin specimens

were collected as follows: the patient was placed in a sitting position, a hot compress was applied and a lid gland massage was performed. Subsequently, a saline-soaked cotton swab was used to gently wipe the lipid secretions draining from the lid gland and cultured with bacteria. Specimens of aqueous humor, vitreous body, lacrimal passage and orbital contents were all taken intraoperatively.

Strain Identification and Antimicrobial Sensitivity Test

Bacterial culture identification was performed in strict accordance with the routine clinical microbiology identification procedures. Specimens were inoculated into bacterial culture medium, fungal culture medium and enrichment culture medium for culture and separation, and then single colonies were selected and identified by Hunan Mindray TDR-300B PLUS microbial identification instrument. The operation of the antimicrobial sensitivity test and the interpretation of the results were carried out in strict accordance with the M100-S20 rules and standards of the Clinical and Laboratory Standards Institute (CLSI)^[26]. The antimicrobial sensitivity test was carried out using the “broth microdilution method” recommended by CLSI to detect the susceptibility and MIC values of the drugs separately. After processing by intelligent expert analysis software, the report results were obtained and reasonable expert evaluation was put forward. The quality control strains were *Staphylococcus aureus* (ATCC25923), *Enterococcus faecalis* (ATCC29212), *Escherichia coli* (ATCC25922) and *Pseudomonas aeruginosa* (ATCC27853).

Statistical Analysis All data in this study were statistically analyzed using SPSS 23.0 software. χ^2 test was utilized to compare categorical variables, with $P < 0.05$ indicating a statistically significant difference.

RESULTS

Distribution Results of Pathogenic Bacteria Among the patients with positive bacterial culture, 84 were male and 71 were female, with a ratio of 1.18:1 and a mean age of 49.65 ± 22.21 y. Of the 155 strains of pathogenic bacteria, gram-positive bacteria predominated at 45.81%, followed by fungi at 39.35%, and gram-negative bacteria accounted for the lowest percentage of 14.84%. *Staphylococcus epidermidis* and *Streptococcus pneumoniae* accounted for the top two gram-positive bacteria infections with 38.06% and 1.94%, respectively. *Pseudomonas aeruginosa* (6.45%) and *Stenotrophomonas maltophilia* (1.94%) were the top two pathogens accounting for the gram-negative bacteria infections, respectively. *Mucor* (20%) and *Fusarium* (15.48%) were the top two pathogens in fungi (Table 1).

Analysis of Specimen Collection Sites and Positive Culture of Specimens The distribution of bacterial infections in different parts of the eye from January 2020 to March 2023

Table 1 Distribution and composition ratio of pathogenic bacteria in patients with eye infections

Pathogenic bacteria	Number	Proportion (%)
Gram-positive bacteria	71	45.81
<i>Staphylococcus epidermidis</i>	59	38.06
<i>Streptococcus pneumoniae</i>	3	1.94
<i>Micrococcus luteus</i>	2	1.29
<i>Staphylococcus intermedius</i>	1	0.65
<i>Staphylococcus haemolyticus</i>	1	0.65
<i>Corynebacterium propinquum</i>	1	0.65
<i>Streptococcus constellatus</i>	1	0.65
<i>Staphylococcus aureus</i>	1	0.65
<i>Aerococcus viridans</i>	1	0.65
<i>Enterococcus faecalis</i>	1	0.65
Gram-negative bacteria	23	14.84
<i>Pseudomonas aeruginosa</i>	10	6.45
<i>Stenotrophomonas maltophilia</i>	3	1.94
<i>Escherichia hermannii</i>	2	1.29
<i>Enterobacter agglomerans</i>	2	1.29
<i>Aeromonas hydrophila</i>	1	0.65
<i>Enterobacter cloacae</i>	1	0.65
<i>Citrobacter koseri</i>	1	0.65
<i>Achromobacter xylooxidans</i>	1	0.65
<i>Serratia marcescens</i>	1	0.65
<i>Klebsiella pneumoniae</i>	1	0.65
Fungi	61	39.35
<i>Mucor</i>	31	20.00
<i>Fusarium</i>	24	15.48
<i>Aspergillus</i>	3	1.94
Others	3	1.94
Total	155	100.00

(Table 2). *Staphylococcus epidermidis* was the most common strain in the conjunctiva, cornea, aqueous humor or vitreous body and other eye parts. Besides, *Fusarium* and *Pseudomonas aeruginosa* were also among the most common strains of conjunctival and corneal infections.

Drug Resistance of Bacterial Strains The results of the antimicrobial sensitivity test showed that gram-positive bacteria were highly resistant to penicillin and erythromycin (78.87% and 46.48% respectively), but least resistant to vancomycin at 0. Gram-negative bacteria were highly resistant to cefoxitin and compound sulfamethoxazole (100% and 95.65% respectively), but least resistant to meropenem at 0 (Table 3). Further comparison of the resistance of gram-negative bacteria and gram-positive bacteria to various drugs revealed statistically significant differences ($P < 0.05$) in the resistance of both to cefoxitin, cotrimoxazole, levofloxacin, cefuroxime, ceftriaxone and ceftazidime, and both had higher rates of resistance to gram-negative bacteria than to gram-positive bacteria (Table 4).

Table 2 Distribution of bacterial infections in different parts of the eye

Bacterial strains isolated from different parts of the eye	Number	Proportion (%)
Cornea	108	69.68
<i>Staphylococcus epidermidis</i>	44	28.39
<i>Fusarium</i>	21	13.55
<i>Mucor</i>	24	15.48
<i>Pseudomonas aeruginosa</i>	4	2.58
<i>Streptococcus pneumoniae</i>	2	1.29
<i>Aspergillus</i>	2	1.29
<i>Escherichia hermannii</i>	2	1.29
<i>Stenotrophomonas maltophilia</i>	1	0.65
<i>Staphylococcus aureus</i>	1	0.65
<i>Micrococcus gluteus</i>	1	0.65
<i>Staphylococcus intermedius</i>	1	0.65
<i>Staphylococcus haemolyticus</i>	1	0.65
<i>Enterobacter cloacae</i>	1	0.65
<i>Citrobacter koseri</i>	1	0.65
<i>Aerococcus viridans</i>	1	0.65
<i>Micrococcus luteus</i>	1	0.65
Dacryocyst	19	12.26
<i>Staphylococcus epidermidis</i>	5	3.23
<i>Fusarium</i>	3	1.94
<i>Pseudomonas aeruginosa</i>	3	1.94
<i>Mucor</i>	2	1.29
<i>Enterobacter agglomerans</i>	2	1.29
<i>Aspergillus</i>	1	0.65
<i>Streptococcus pneumoniae</i>	1	0.65
<i>Stenotrophomonas maltophilia</i>	1	0.65
<i>Enterococcus faecalis</i>	1	0.65
Aqueous humor or vitreous body	17	10.97
<i>Staphylococcus epidermidis</i>	5	3.23
<i>Mucor</i>	5	3.23
<i>Pseudomonas aeruginosa</i>	3	1.94
<i>Stenotrophomonas maltophilia</i>	1	0.65
<i>Corynebacterium propinquum</i>	1	0.65
<i>Streptococcus constellatus</i>	1	0.65
<i>Aeromonas hydrophila</i>	1	0.65
Other eye parts	11	7.10
<i>Staphylococcus epidermidis</i>	5	3.23
<i>Achromobacter xylooxidans</i>	1	0.65
<i>Serratia marcescens</i>	1	0.65
<i>Klebsiella pneumoniae</i>	1	0.65
Others	3	1.94
Total	155	100

DISCUSSION

The eyes, as a vital visual structure in people's lives, are the basis for normal visual function in a healthy state. Infections in ophthalmic patients may easily cause vision loss and affect

Table 3 Drug resistance of bacterial strains

Drugs	Number of drug-resistant patients
Gram-positive bacteria (n=71)	
Penicillin	56
Erythromycin	33
Azithromycin	28
Oxacillin	27
Cefoxitin	24
Compound sulfamethoxazole	24
Levofloxacin	20
Cefuroxime	18
Clarithromycin	17
Clindamycin	16
Tetracycline	16
Ceftriaxone	14
Ceftazidime	11
Vancomycin	0
Gram-negative bacteria (n=23)	
Cefoxitin	23
Compound sulfamethoxazole	22
Levofloxacin	18
Cefuroxime	15
Ceftriaxone	10
Ceftazidime	10
Gentamicin	6
Minocycline	4
Cefotaxime	3
Ampicillin	3
Linezolid	1
Aztreonam	1
Amikacin	1
Meropenem	0

the effectiveness of treatment. Therefore, in case of persistent ocular discomfort, medical attention should be sought as soon as possible and ophthalmic antimicrobial drugs should be used appropriately to treat infectious ophthalmic diseases thoroughly and effectively and to improve their treatment outcome^[27-29]. Farmers, construction workers and other groups are more prone to infection among the many people with eye infections because of their frequent farming and labor. Specifically, farmers are easily scratched by plants and splashed with seeds during labor, which leads to fungal infection; Construction workers are more likely to be infected with gram-positive bacteria because they are often hooked by wires or hit by stones. For this reason, protective eyewear should be used properly during labor to reduce the incidence of eye infections. Supplemented by the increasing use of electronic products in the context of the rapid changes in technology, eye fatigue and reduced eye immunity are becoming more common,

Table 4 Comparison of resistance of gram-positive and gram-negative bacteria to some drugs

Items	Cefoxitin	Compound sulfamethoxazole	Levofloxacin	Cefuroxime	Ceftriaxone	Ceftazidime
Gram-positive bacteria (n=71)						
Number of drug-resistant plants	24	24	20	18	14	11
Proportion	33.80%	33.80%	28.17%	25.35%	19.72%	15.49%
Gram-negative bacteria (n=23)						
Number of drug-resistant plants	23	22	18	15	10	10
Proportion	100%	95.65%	78.26%	65.22%	43.48%	43.48%
χ^2	30.451	26.594	18.100	12.119	5.158	7.842
P	<0.001	<0.001	<0.001	<0.001	0.023	0.005

eventually leading to eye infections. Meanwhile, people's pursuit of beauty has improved under the rising standard of living, contributing to a rapid increase in the use of contacts lenses and cosmetic contact lenses. But with this comes their unregulated use, causing adverse eye consequences such as eye dryness, fatigue and discomfort, and eye infections^[30].

Bacterial infections, as the most common pathogenic bacteria in eye infections, are constantly changing due to the continuous application and even abuse of antimicrobial drugs and hormones, resulting in changing drug resistance as well^[31]. Early identification of pathogenic bacteria species as well as a timely and accurate selection of antimicrobial drugs are key to treating patients with eye infections^[32-33]. In this study, the clinical data of 155 patients admitted to Lixiang Eye Hospital of Soochow University in Suzhou from January 2020 to March 2023 were retrospectively collected, and the pathogenic bacteria species and drug resistance of each pathogenic bacteria were analyzed. The results showed that the pathogens infecting patients with eye infections were mainly gram-positive bacteria, followed by fungi and gram-negative bacteria. The main gram-positive bacteria were *Staphylococcus epidermidis* (45.04%) and *Streptococcus pneumoniae* (2.29%), whereas the main gram-negative bacteria were *Pseudomonas aeruginosa* (7.63%) and *Stenotrophomonas maltophilia* (2.29%), which was consistent with the predominance of gram-positive bacteria in eye infections reported in related studies^[34]. The analysis of the infected site shows that the most infected site was the conjunctiva (50.38%), followed by the cornea (31.30%) and finally the aqueous humor or vitreous body (9.92%).

In previous reports, no bacteria isolated from all types of eye infections were found to be resistant to vancomycin and the sensitivity of the drug was definitive^[35-36]. Comparison of the resistance of gram-positive and gram-negative bacteria to some drugs revealed that gram-positive bacteria were highly resistant to penicillin and erythromycin (78.87% and 46.48% respectively), but least resistant to vancomycin at 0 and that gram-negative bacteria were highly resistant to cefoxitin and compound sulfamethoxazole (100% and 95.65% respectively), but least resistant to meropenem at 0. Therefore,

patients diagnosed with gram-positive bacteria can be treated with vancomycin, and treatment with ceftazidime is equally effective; while those diagnosed with gram-negative bacteria are recommended to be treated with meropenem, amikacin, and aztreonam. Further comparison of the resistance of gram-positive bacteria and gram-negative bacteria to some drugs revealed statistically significant differences ($P < 0.05$) in the resistance of both to cefoxitin, compound sulfamethoxazole, levofloxacin, cefuroxime, ceftriaxone and ceftazidime, and both had higher rates of resistance to gram-negative bacteria than to gram-positive bacteria. This suggests that the above drugs are less effective in gram-negative bacterial infections, and can be used in those with gram-positive bacterial infections. Considering that the resistance of pathogenic bacteria may be attributed to improper or abusive use of previous antimicrobial drugs, there is a need to improve the norms and systems of drug use in the hospital and to provide targeted treatment for postoperative patients with different pathogenic bacterial infections, so as to achieve the best therapeutic effect^[37].

Based on the results of this study, ophthalmologists may make more appropriate decisions in the treatment of eye-bacterial infections. Nevertheless, certain limitations are also visible in the present study. First, this study was a single-center study with accidental results, which led to differences in the distribution of pathogenic bacteria in this paper compared with other literature. Second, a small sample size was included in this study, resulting in low test efficiency and incomplete results. Therefore, the sample size should be further increased in subsequent studies. Finally, the retrospective approach to the study and the time available for the study are limited due to the nature of the methods, which limits our ability to perform a more objective analysis of trends in pathogen distribution and antibiotic sensitivity. To address this, studies of eye bacterial infections with larger sample sizes, longer duration, and using more advanced technology will be conducted in the future.

To put it in a nutshell, *Staphylococcus epidermidis* is the main isolated strain of all eye bacterial specimens in Suzhou, China. Antibiotics such as penicillin and erythromycin, used

for the treatment of gram-positive bacterial infections, are less effective, and vancomycin is the best, followed by ceftazidime. Cefoxitin and compound sulfamethoxazole are less effective in the treatment of gram-negative bacterial infections, so meropenem is recommended. By further classifying bacteria and comparing their sensitivity to antibiotics through statistical analysis, more accurate results of antibiotic sensitivity analysis can be obtained.

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