

Long term follow-up of a family with *GUCY2D* dominant cone dystrophy

Georgios Tsokolas^{1,2}, Hussein Almuhtaseb^{1,2}, Helen Griffiths¹, Fatima Shawkat², Reuben J. Pengelly³, Sarah Ennis³, Andrew Lotery^{1,2}

¹Clinical and Experimental Sciences, University of Southampton, Southampton, Hampshire SO16 6YD, UK

²Eye Unit, University Hospital Southampton, Southampton, Hampshire SO16 6YD, UK

³Human Genetics & Genomic Medicine, Faculty of Medicine, University of Southampton, Southampton, Hampshire, SO16 6YD, UK

Correspondence to: Andrew Lotery. Clinical and Experimental Sciences, University of Southampton, Southampton, Hampshire SO16 6YD, UK. a.j.lotery@soton.ac.uk

Received: 2018-05-15 Accepted: 2018-09-06

Abstract

• **AIM:** To describe long term follow-up in a family with *GUCY2D* dominant cone dystrophy.

• **METHODS:** Optical coherence tomography scans and fundus autofluorescence images were obtained. Flash and pattern electroretinograms (ERGs) and occipital pattern reversal visual evoked potentials were recorded.

• **RESULTS:** Two members of the same family (father and son) were identified to have the heterozygous R838C mutation in the *GUCY2D* gene. The father presented at the age of 45 with bilateral bull's eye maculopathy and temporal disc pallor. Over 13y of serial follow up visits, the bull's eye maculopathy progressed gradually into macular atrophy. Electrophysiological tests were significantly degraded suggesting poor macular function. Spectral-domain optical coherence tomography (SD-OCT) scans showed progressive loss and disruption of the ellipsoid layer at the foveal level. His son presented at the age of 16 with bilateral granular retinal pigment epithelial changes in both maculae. Electrophysiological testing was initially borderline normal but has gradually deteriorated to show reduced cone ERGs and macula function. SD-OCT demonstrated gradual macular thinning and atrophy bilaterally. Unlike his father, there was no disruption of the ellipsoid layer.

• **CONCLUSION:** Both family members exhibited gradual changes in their fundi, electrophysiological testing and multimodal imaging. Changes were milder than those observed in other mutations of the same gene.

• **KEYWORDS:** autofluorescence; electroretinogram; cone dystrophy; cone-rod dystrophy; *GUCY2D*; spectral-domain optical coherence tomography; visual evoked potential

DOI:10.18240/ijo.2018.12.12

Citation: Tsokolas G, Almuhtaseb H, Griffiths H, Shawkat F, Pengelly RJ, Sarah E, Lotery A. Long term follow-up of a family with *GUCY2D* dominant cone dystrophy. *Int J Ophthalmol* 2018;11(12):1945-1950

INTRODUCTION

Cone dystrophies (CD) and cone-rod dystrophies (CRD) are a group of genetic disorders, which demonstrate a large degree of heterogeneity and severity. The most frequent mode of inheritance is autosomal dominant, although autosomal recessive and X-linked recessive modes of inheritance have also been reported^[1]. The main symptoms are decreased central visual acuity (VA), markedly decreased color vision, hemeralopia, nystagmus and loss of peripheral vision^[2]. In pure cone dystrophies, only cone function is affected, while rod function remains intact. The photopic electroretinograms (ERG) demonstrates abnormalities but the scotopic ERG is grossly normal. Conversely, in cone-rod dystrophies, patients demonstrate features suggestive of rod dysfunction as well. In CRD, both photopic and scotopic ERGs will be abnormal^[1-3]. It is rare to have a pure cone dystrophy because of the reciprocal relationship between the cone and rod system^[3].

The usual natural history of CRD starts initially by forming some non-specific retinal pigment epithelial (RPE) granularity and mottling at the macular level. As the disease progresses, a typical bull's eye lesion develops, but not universally. End-stage disease with photoreceptor degeneration and RPE loss will result in geographic atrophy^[3]. There is a wide range of genes implicated in the pathogenesis of CRD. The most common ones are *SEMA4A*, *AIPL1*, *CRX*, *GUCA1A*, *GUCY2D*, *PITPNM3*, *PRPH2*, *PROM1*, *RIMS1* and *UNC119*^[4].

Both family members were found to have mutation in the retinal guanylyl cyclase 1 gene (also known as guanylate cyclase 2D/*GUCY2D*), which is known to be implicated in the autosomal dominant form of cone/cone-rod dystrophy. Retinal guanylyl cyclase 1 is an enzyme expressed within the retina responsible for the conversion of guanosine 5'-triphosphate

Clinical features in *GUCY2D* dominant cone dystrophy

to cyclic guanosine monophosphate (cGMP)^[4]. Like other membrane guanylyl cyclases, this enzyme has a hydrophobic amino-terminal signal sequence followed by a large extracellular domain, a single membrane spanning domain, a kinase homology domain, and a guanylyl cyclase catalytic domain. In contrast to other membrane guanylyl cyclases, this enzyme is not activated by natriuretic peptides. Retinal guanylyl cyclase 1 helps photoreceptors return to their dark-adapted state after light exposure; cGMP plays a significant role as the second messenger molecule in the phototransduction cascade by keeping the voltage-gated sodium and calcium channels of photoreceptors open. Photoactivation leads to conversion of cGMP to guanosine 5'-monophosphate by phosphodiesterase and this results in the closure of voltage-gated sodium and calcium channels and to hyperpolarization of the photoreceptor outer segments. When the concentration of calcium cations is reduced, retinal guanylyl cyclase 1 restores the levels of cGMP and this allows the reopening of the relevant channels. Restoration of cGMP levels is achieved by the presence of the guanylate cyclase-activating protein^[4]. Mutations in *GUCY2D* gene have been described in cone-rod dystrophy-6 and Leber congenital amaurosis^[5].

In this manuscript, we describe the long-term clinical and multimodal imaging findings over the course of 13y in two family members diagnosed with *GUCY2D* cone dystrophy. To the best of our knowledge, this is the longest follow-up described so far in literature.

SUBJECTS AND METHODS

All procedures were compliant and consistent with the tenets of the Declaration of Helsinki. Informed consent was obtained from all individual participants included in this retrospective study. A retrospective review of the electronic records of two family members (father and son) was conducted at the Eye Unit of University Hospital Southampton National Healthcare System Foundation Trust, UK. Both patients were followed-up annually at the Eye Unit for the last 13y and had a full past medical, ophthalmic and genetic history taken during the initial presentation. Annual follow-up visits were conducted including multimodal imaging. Optical coherence tomography (OCT) scans were obtained with the use of Triton/OCT-2000 (Topcon Ltd, Tokyo, Japan), whereas fundus autofluorescence (FAF) images were taken using Spectralis (Heidelberg Engineering, Heidelberg, Germany). Goldmann visual field (GVF) testing was utilised to monitor the progression of central field loss. Flash and pattern ERGs were recorded using corneal Dawson-Trick-Litzkow (DTL) thread electrodes. Flash ERGs were recorded after dilatation in compliance with International Society for Clinical Electrophysiology of Vision standards^[6]. Occipital full-field checkerboard reversal visual evoked potentials (VEPs) were recorded to stimulus check sizes ranging from 10 to 120min of arc.

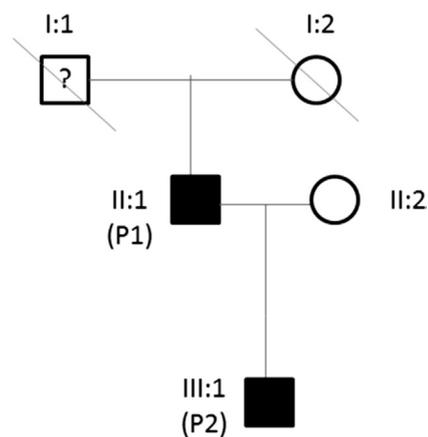


Figure 1 Family pedigree.

Both patients underwent genetic testing: the participants underwent whole exome sequencing in order to identify the genetic cause of their CD. DNA was isolated from blood, exome enrichment performed using the Agilent SureSelect Human All Exon V5 kit (© Agilent Technologies, Inc), and sequencing performed on the Illumina HiSeq 2000 platform (© Illumina Inc®). Data analysis was performed as previously described^[7]. Genetics variants were filtered to identify variants present in both individuals within candidate genes identified through the Human Gene Mutation Database (namely *ABCA4*, *CACNA2D4*, *CNGA3*, *CNGB3*, *CRB1*, *CRX*, *GUCA1A*, *GUCY2D*, *KCNV2*, *MERTK*, *orf15*, *PDE6C*, *PDE6H*, *PITPNM3* & *PRPH2*).

RESULTS

Genetic Testing Exome sequencing identified the heterozygous variant *GUCY2D*:c.2512C>T:p.Arg838Cys (rs61750172, also known as R838C) in both patients (Figure 1). This mutation has been previously reported to cause cone-rod dystrophy 6 (CORD6; OMIM #601777)^[8]. For abbreviation purposes, the father has been allocated the symbol P1 in generation II, whereas the son has been allocated the symbol P2 in generation III. There was also a history of eye problems in patient II. There was insufficient data in past medical history to confirm a formal diagnosis of CD; hence the question mark symbol (Figure 1).

Clinical Findings The cumulative clinical features for each patient are summarized in Table 1. Both patients exhibited decline in VA combined with hemeralopia in adolescence but neither of them complained of nyctalopia. In addition, there were no significant media opacities to account for decline in VA in both of our patients. Figure 2 demonstrates the fluctuation of best-corrected visual acuity (BCVA) in both family members over the 13y follow-up at Southampton Eye Unit.

Multimodal Imaging The macular OCT scans of P1 showed progressive loss of the ellipsoid layer at the level of the fovea with gradual thinning and atrophy of the adjacent retinal tissue and reverse shadowing due to cone and RPE cell loss

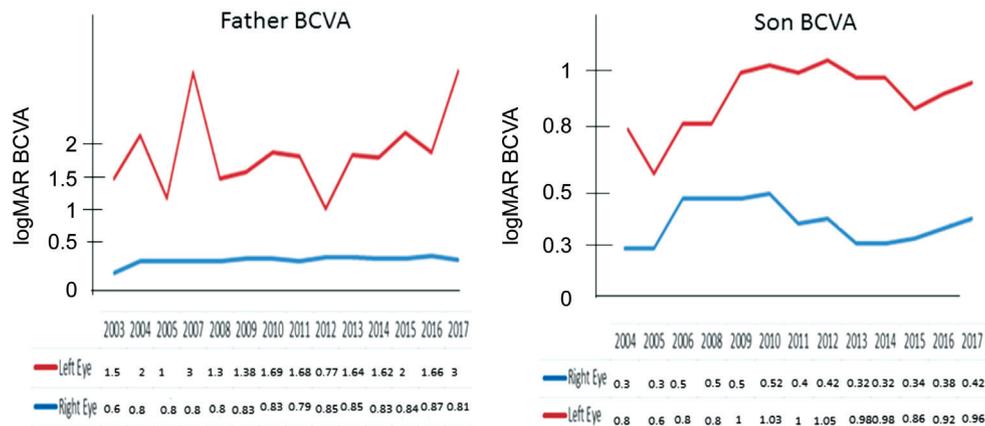


Figure 2 Changes in BCVA for both patients from initial presentation in 2004 until 2017.

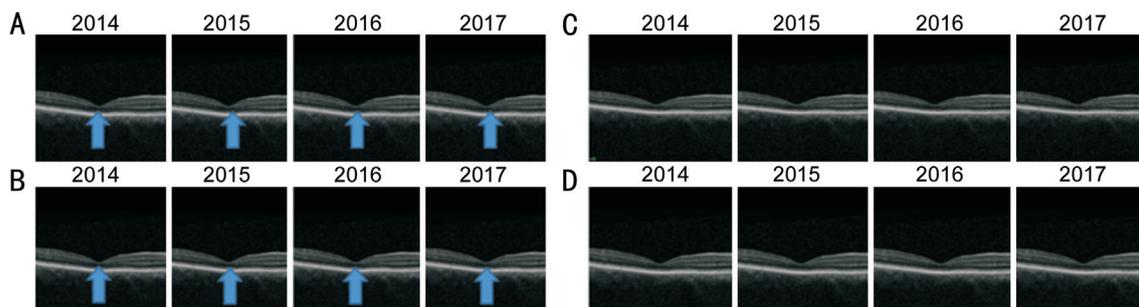


Figure 3 Serial OCT images both patients A: P1's right eye; B: P1's left eye; C: P2's right eye; D: P2's left eye. Progressive atrophy of the ellipsoid layer but no breaks in the continuity of the ellipsoid layers on the OCT images.

Table 1 Cumulative table summarizing the clinical features in both family members

| Pedigree | Current age (y) | Onset of symptoms | Ocular comorbidities | VA (Snellen) OD, OS | Ishihara plates | Dilated fundal examination findings | GVFs | ERG/VEP findings |
|-------------|-----------------|----------------------------------|---|----------------------|----------------------|---|-----------------------------|---|
| II: 2 (P1) | 59 | Photophobia since adolescence | Left eye amblyopia due to squint | 6/36, hand movements | OD: 1/17 OS: 0/17 | Bilateral bull's eye maculopathy and temporal disc pallor | Progressive central scotoma | Impaired cone function, preserved rod function, degraded and attenuated VEP |
| III: 3 (P2) | 28 | Photophobia in early adolescence | Bilateral astigmatism, left eye amblyopia | 6/15, 6/48 | OD: 5/17 OS: 1/17 | Bilateral RPE changes | Progressive central scotoma | Impaired cone function, preserved rod function, degraded and attenuated VEP |

VA: Visual acuity; GVFs: Goldmann visual fields; ERG: Electroretinograms; VEP: Visual evoked potentials. Neither of the affected family members reported symptoms of nyctalopia or had significant cataracts.

(Figure 3A, 3B). FAF showed a central annular area of hypo-autofluorescence corresponding to macular atrophy and RPE loss with a surrounding ring of hyper-autofluorescence (hyper-AF) indicating the transition zone between normal and abnormal retina (Figure 4A, 4B). These changes have occurred in both eyes but the left eye appears to be more affected than the right. The macular OCT scan of the right eye of P2 demonstrated gradual macular thinning and atrophy, whereas the macular structure in the left eye remained relatively stable. Unlike the father's OCT scans, there was no disruption of the ellipsoid layer (Figure 3C, 3D). FAF images of the son showed features suggestive of bilateral foveolar hyper-AF. The hyper-AF involving the central foveolar area which can be seen in Figure 4C and 4D, are similar to the changes previously reported in type-2 idiopathic macular telangiectasia^[9]. FAF findings also demonstrated RPE granular changes.

Electrophysiology The father's flash ERGs showed well-preserved rod function (amplitude of responses smaller than average but within normal range) but significantly impaired cone function. Pattern ERGs as well occipital pattern VEPs were attenuated and degraded indicating reduced macular function. The son's cone responses were of borderline normal amplitude on initial presentation but became significantly degraded ten months later suggesting cone dysfunction (Figure 5). Rod responses were normal. Pattern ERGs and occipital pattern VEPs were significantly degraded indicating reduced macular function.

DISCUSSION

So far, 223 mutations in the *GUCY2D* gene have been described. The *Arg838Cys (R838C)* mutation described in both of our patients has been previously reported by Kellsell *et al*^[10] in 1998 in cone dystrophy 6. It is reported to cause

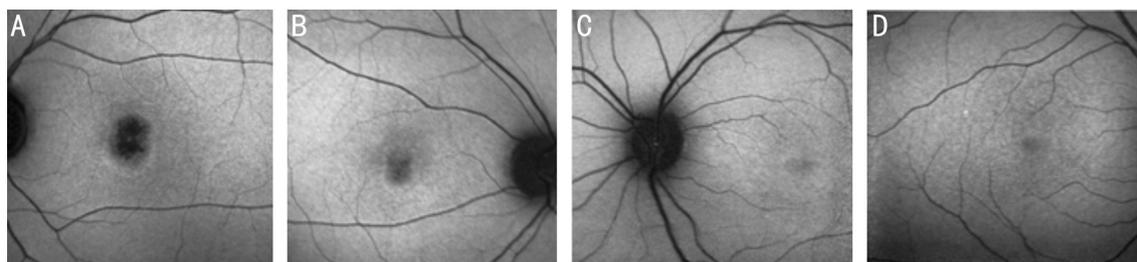


Figure 4 Autofluorescence images from both patients A, B: Father (P1): central area of hypo-autofluorescence, left worse than right; C, D: Son (P2): bilateral foveolar hyper-AF. The hyper-AF changes may be a consequence of decreased foveal pigment density and secondarily reduced masking effect of the RPE fluorescence.

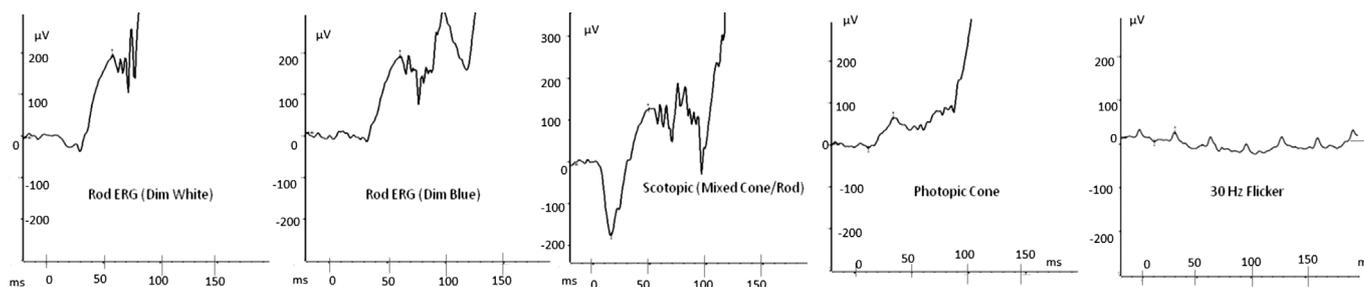


Figure 5 P2's ERG responses It shows P2's repeated ERG responses a few months after initial presentation. There was a significant reduction in the amplitude of the cone mediated responses, which were more degraded compared to the initial ERGs.

a milder clinical phenotype compared to other mutations in the *GUCY2D* gene^[11]. The mild phenotype of this particular mutation has been described by others in the past^[12-13]. Other *GUCY2D* mutations on the same codon (R838S, R838H, R838P, R838G) can lead to a more aggressive clinical picture^[14-15]. Based on ERG recordings, two major types of cone-rod dystrophy were differentiated according to the phenotypic classification by Szlyk *et al*^[16]. In type 1, cone amplitudes were reduced to a greater degree than rod amplitudes, while in type 2, cone and rod ERG amplitudes were reduced in equal proportion. According to the phenotypic classification by Szlyk *et al*^[16], both of our patients could be classified as phenotype 1a.

The father presented to the Ophthalmology Department with bilateral bull's eye maculopathy and mild temporal disc pallor. Bull's eye maculopathy can be caused by genetically inherited conditions or toxic retinopathies, hence it is not disease specific for cone/cone-rod dystrophy^[3]. Disc pallor is also a non-specific finding but has been reported previously in a patient with cone dystrophy^[17], who had normal to near normal VA and color vision and abnormal peripheral cone function. However, in the father's case, the macula function was already compromised and the peripheral retina was normal. His son exhibited non-specific RPE granular changes but no other significant abnormalities. Bull's eye maculopathy was not observed in the son's case confirming that bull's eye maculopathy is not a universal sign^[3]. This may be merely due to the chronicity of the disorder in his father. Moreover, the presence of a bull's eye maculopathy does not always correlate

accurately with the extent of retinal dysfunction^[18]. All the above observations confirm that the diagnosis of CD/CRD cannot rely exclusively on funduscopy due to the non-specific clinical findings^[19].

On spectral-domain optical coherence tomography (SD-OCT), the father exhibited progressive loss of the ellipsoid layer and gradual thinning and atrophy of the parafoveal retinal tissue and reverse shadowing due to cone and RPE cell loss. There was also obscurity at the level of the external limiting membrane (EML). This is consistent with the findings of others^[19-23]. His son, however, had thinning but no loss of the ellipsoid layer. FAF imaging from the father's fundus was consistent with the OCT findings. Furthermore, the surrounding hyper-AF around the annular area of hypo-autofluorescence suggests gradual deposition of lipofuscin material, a byproduct of the photoreceptor cell visual cycle and RPE metabolism. Lipofuscin accumulation can be toxic to the RPE and photoreceptor cells and this can lead to death of RPE and photoreceptors and that can cause further thinning and atrophy of the macula^[24]. The FAF findings from the father's fundus are consistent with observations of another paper^[25]. The RPE granular changes observed in the son's fundus were also observed by FAF. Hence, FAF is useful as an adjuvant means of imaging when SD-OCT cannot detect subtle RPE or retinal abnormalities. FAF images showing a subtle bilateral hyper-AF signal mimicking changes that were previously described in Type 2 Macular Telangiectasia might be a reliable early indicator of the disease especially when ERGs are found to be border-line normal as in P2 in this case series^[7].

Only one paper by Cho *et al*^[26] has attempted to describe in depth and classify the different types of structural retinal abnormalities in patients with CD/CRD. This was a five year observational follow-up in 15 patients with cone dystrophy. Prior to this study, Hood *et al*^[21] reported decreased intensity in the ellipsoid layer in 6 patients with cone dystrophy. Birch *et al*^[27] reported that the thickness of the outer nuclear layer and the sum of thickness of the RPE and outer segment correlated well with visual field sensitivity^[20,24]. However, neither paper described the structural changes of the retina in patients with cone dystrophies.

Cho *et al*^[26] divided the morphological changes in the retinal structure in cone dystrophy patients into four different categories: 0, 1, 2 and 3. Category 0 exhibited no structural abnormalities, whereas category 1 showed foveal ellipsoid layer loss and obscuring of the border between the ellipsoid band and ELM. Category 2 showed foveal thinning and focal foveal ellipsoid layer disruption with an intact ELM. Finally, category 3 showed foveal thickening and perifoveal disruption of the ellipsoid layer.

Based on this classification, the father demonstrated changes matching category 1. The son did have foveal thinning but no disruption of the ellipsoid layer, hence he could potentially be classified as category 0. In the Cho *et al*'s^[26] paper, it was observed that category 0 patients were younger than the other categories, although this observation was not proven to be statistically significant. Ageing is likely to be a contributing factor to disease progression with subsequent disruption of the photoreceptor outer segment/ellipsoid layer.

Moreover, in the paper by Cho *et al*^[26], only one patient was found to meet the category 3 criteria. The authors formed the hypothesis that the thickening of the fovea could be attributed to the gradual deposition of tissue remnants of the unhealthy and gradually dying photoreceptors. This SD-OCT finding has been described previously in patients with *peripherin/RDS* gene mutations^[28]. However, in patients with CD due to *GUCY2D* mutations, category 1 abnormalities have been previously described (*GUCY2D Arg838His*). The difference to the case described by Kim *et al*^[29] (*GUCY2D Arg838His*) is that our patients had the *GUCY2D Arg838Cys* mutation. Nevertheless, the morphological features on SD-OCT are similar.

Electrophysiological testing is arguably the most diagnostic test, should CD/CRD be suspected. Fundus examination can show non-specific changes and the multimodal imaging findings in patients with cone dystrophy are quite heterogeneous and therefore funduscopy, SD-OCT and FAF are not diagnostic. This is also supported by Cho *et al*^[26], who observed that category 0 patients had a significantly affected ERG, while no structural abnormalities were observed. In addition, it is obvious that electrophysiological responses do not correlate

well with SD-OCT findings. Thus, ERG and VEP can confirm the macula/cone dysfunction much earlier than other imaging modalities and therefore they both are an irreplaceable adjuvant diagnostic tool for any Medical Retina Specialist in the diagnosis and management of patients diagnosed with CD/CRD.

To the best of our knowledge, this is the longest duration of follow-up of patients with CD associated with mutations in *GUCY2D*. We describe the progression of the disease based on VA and multimodal imaging. Electrophysiological testing is most useful for the clinical diagnosis of CD/CRD, while SD-OCT and FAF imaging are both useful for monitoring disease progression and genotype-phenotype correlations can be identified by molecular analysis.

ACKNOWLEDGEMENTS

Authors would like to express gratitude to the patients for their involvement in this study. We thank Dr. David Bunyan, Senior Clinical Scientist, Salisbury National Healthcare System Foundation Trust for confirmatory genetic testing and Ms. Angela Cree, Senior Research Manager, University of Southampton for overall project coordination. We thank Fight against Blindness Charity Organization for supporting this project.

Foundation: Supported by Fight Against Blindness Charity Organization.

Conflicts of Interest: Tsokolas G, None; Almuhtaseb H, None; Griffiths H, None; Shawkat F, None; Pengelly RJ, None; Ennis S, None; Lotery A, None.

REFERENCES

- 1 Aboshiha J, Dubis AM, Carroll J, Hardcastle AJ, Michaelides M. The cone dysfunction syndromes. *Br J Ophthalmol* 2016;100(1):115-121.
- 2 Thiadens AA, Phan TM, Zekveld-Vroon RC, *et al*. Clinical course, genetic etiology, and visual outcome in cone and cone-rod dystrophy. *Ophthalmology* 2012;119(4):819-826.
- 3 Kanski J. *Clinical ophthalmology, a systematic approach*. Elsevier 2015.
- 4 Nong E, Lee W, Merriam JE, Allikmets R, Tsang SH. Disease progression in autosomal dominant cone-rod dystrophy caused by a novel mutation (D100G) in the *GUCA1A* gene. *Doc Ophthalmol* 2014;128(1):59-67.
- 5 Boye SE. Leber congenital amaurosis caused by mutations in *GUCY2D*. *Cold Spring Harb Perspect Med* 2014;5(1):a017350.
- 6 McCulloch DL, Marmor MF, Brigell MG, Hamilton R, Holder GE, Tzekov R, Bach M. ISCEV standard for full-field clinical electroretinography (2015 update). *Doc Ophthalmol* 2015;130(1):1-12.
- 7 Pengelly RJ, Upstill-Goddard R, Arias L, Martinez J, Gibson J, Knut M, Collins AL, Ennis S, Collins A, Briceno I. Resolving clinical diagnoses for syndromic cleft lip and/or palate phenotypes using whole-exome sequencing. *Clin Genet* 2015;88(5):441-449.
- 8 Jiang F, Xu K, Zhang X, Xie Y, Bai F, Li Y. *GUCY2D* mutations in a Chinese cohort with autosomal dominant cone or cone-rod dystrophies. *Doc Ophthalmol* 2015;131(2):105-114.

Clinical features in *GUCY2D* dominant cone dystrophy

- 9 Toto L, Di Antonio L, Mastropasqua R, Mattei PA, Carpineto P, Borrelli E, Rispoli M, Lumbroso B, Mastropasqua L. Multimodal imaging of macular telangiectasia type 2: focus on vascular changes using optical coherence tomography angiography. *Invest Ophthalmol Vis Sci* 2016;57(9):OCT268-276.
- 10 Kelsell RE, Gregory-Evans K, Payne AM, Perrault I, Kaplan J, Yang RB, Garbers DL, Bird AC, Moore AT, Hunt DM. Mutations in the retinal guanylate cyclase (RETGC-1) gene in dominant cone-rod dystrophy. *Hum Mol Genet* 1998;7(7):1179-1184.
- 11 Wilkie SE, Newbold RJ, Deery E, Walker CE, Stinton I, Ramamurthy J, Hurley JB, Bhattacharya SS, Warren MJ, Hunt DM. Functional characterization of missense mutations at codon 838 of retinal guanylate cyclase correlates with disease severity in patients with autosomal dominant cone-rod dystrophy. *Hum Mol Genet* 2000;9(20):3065-3073.
- 12 Downes SM, Payne AM, Kelsell RE, Fitzke FW, Holder GE, Hunt DM, Moore AT, Bird AC. Autosomal dominant cone-rod dystrophy with mutations in the guanylate cyclase 2D gene encoding retinal guanylate cyclase-1. *Arch Ophthalmol* 2001;119(11):1667-1673.
- 13 Van Ghelue M, Eriksen HL, Ponjavic V, Fagerheim T, Andreasson S, Forsman-Semb K, Sandgren O, Holmgren G, Tranebjaerg L. Autosomal dominant cone-rod dystrophy due to a missense mutation (R838C) in the guanylate cyclase 2D gene (*GUCY2D*) with preserved rod function in one branch of the family. *Ophthalmic Genet* 2000;21(4):197-209.
- 14 Gregory-Evans K, Kelsell RE, Gregory-Evans CY, Downes SM, Fitzke FW, Holder GE, Simunovic M, Mollon JD, Taylor R, Hunt DM, Bird AC, Moore AT. Autosomal dominant cone-rod retinal dystrophy (CORD6) from heterozygous mutation of *GUCY2D*, which encodes retinal guanylate cyclase. *Ophthalmology* 2000;107(1):55-61.
- 15 Mukherjee R, Robson AG, Holder GE, Stockman A, Egan CA, Moore AT, Webster AR. A detailed phenotypic description of autosomal dominant cone dystrophy due to a de novo mutation in the *GUCY2D* gene. *Eye (Lond)* 2014;28(4):481-487.
- 16 Szlyk JP, Fishman GA, Alexander KR, Peachey NS, Derlacki DJ. Clinical subtypes of cone-rod dystrophy. *Arch Ophthalmol* 1993;111(6):781-788.
- 17 Ito N, Kameya S, Gocho K, Hayashi T, Kikuchi S, Katagiri S, Gekka T, Yamaki K, Takahashi H, Tsuneoka H. Multimodal imaging of a case of peripheral cone dystrophy. *Doc Ophthalmol* 2015;130(3):241-251.
- 18 Kurz-Levin MM, Halfyard AS, Bunce C, Bird AC, Holder GE. Clinical variations in assessment of bull's-eye maculopathy. *Arch Ophthalmol* 2002;120(5):567-575.
- 19 Kellner U, Kellner S. Clinical findings and diagnostics of cone dystrophy. *Ophthalmologe* 2009;106(2):99-108.
- 20 Mallapatna A, Vinekar A, Jayadev C, Dabir S, Sivakumar M, Krishnan N, Mehta P, Berendschot T, Yadav NK. The use of handheld spectral domain optical coherence tomography in pediatric ophthalmology practice: our experience of 975 infants and children. *Indian J Ophthalmol* 2015;63(7):586-593.
- 21 Hood DC, Zhang X, Ramachandran R, Talamini CL, Raza A, Greenberg JP, Sherman J, Tsang SH, Birch DG. The inner segment/outer segment border seen on optical coherence tomography is less intense in patients with diminished cone function. *Invest Ophthalmol Vis Sci* 2011;52(13):9703-9709.
- 22 Thomas MG, Kumar A, Kohl S, Proudlock FA, Gottlob I. High-resolution in vivo imaging in achromatopsia. *Ophthalmology* 2011;118(5):882-887.
- 23 Leng T, Marmor MF, Kellner U, Thompson DA, Renner AB, Moore W, Sowden JC. Foveal cavitation as an optical coherence tomography finding in central cone dysfunction. *Retina* 2012;32(7):1411-1419.
- 24 Yung M, Klufas MA, Sarraf D. Clinical applications of fundus autofluorescence in retinal disease. *Int J Retina Vitreous* 2016;2:12.
- 25 Wang NK, Chou CL, Lima LH, Cella W, Tosi J, Yannuzzi LA, Tsang SH. Fundus autofluorescence in cone dystrophy. *Doc Ophthalmol* 2009;119(2):141-144.
- 26 Cho SC, Woo SJ, Park KH, Hwang JM. Morphologic characteristics of the outer retina in cone dystrophy on spectral-domain optical coherence tomography. *Korean J Ophthalmol* 2013;27(1):19-27.
- 27 Birch DG, Wen Y, Locke K, Hood DC. Rod sensitivity, cone sensitivity, and photoreceptor layer thickness in retinal degenerative diseases. *Invest Ophthalmol Vis Sci* 2011;52(10):7141-7147.
- 28 Duncan JL, Talcott KE, Ratnam K, Sundquist SM, Lucero AS, Day S, Zhang Y, Roorda A. Cone structure in retinal degeneration associated with mutations in the peripherin/RDS gene. *Invest Ophthalmol Vis Sci* 2011;52(3):1557-1566.
- 29 Kim BJ, Ibrahim MA, Goldberg MF. Use of spectral domain OCT to visualize photoreceptor abnormalities in cone/rod dystrophy-6. *Retin Cases Brief Rep* 2011;5(1):56-61.